

# *Glucosinolates, myrosinase hydrolysis products, and flavonols found in rocket (Eruca sativa and Diplotaxis tenuifolia)*

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## Glucosinolates, myrosinase hydrolysis products and flavonols found in rocket (Eruca

## sativa and Diplotaxis tenuifolia)

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#### 1 Abstract

Rocket species have been shown to have very high concentrations of glucosinolates and flavonols, which have numerous positive health benefits with regular consumption. In this review we highlight how breeders and processors of rocket species can utilize genomic and phytochemical research to improve varieties and enhance the nutritive benefits to consumers. Plant breeders are increasingly looking to new technologies such as HPLC, UPLC, LC-MS and GC-MS to screen populations for their phytochemical content to inform plant selections. Here we collate the research that has been conducted to-date in rocket, and summarise all glucosinolate and flavonol compounds identified in the species. We emphasize the importance of the broad screening of populations for phytochemicals and myrosinase degradation products, as well as unique traits that may be found in underutilized gene bank resources. We also stress that collaboration with industrial partners is becoming essential for long-term plant breeding goals through research.

14 Key words: Brassicaceae, Isothiocyanates, Plant breeding, Indoles, Nitriles

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#### 26 Introduction

27 In recent years, several species of minor leafy-crops have risen to prominence as potentially 28 important commercial and edible species. One example is rocket, which has quickly gained 29 popularity in the Western diet. Originally found as an obscure crop in Mediterranean and 30 Middle-Eastern countries, rocket has become popular largely due to the pungent aromas and 31 tastes associated with it. Glucosinolates (GSLs)/isothiocyanates (ITCs) and flavonols derived from many species <sup>1-4</sup> have been shown to infer significant protection against cancer and 32 heart disease <sup>4–11</sup>. In Western countries, diets are generally lacking in fruits and vegetables. 33 34 Despite government initiatives (such as the "5-a-day" campaign in the UK and USA), these 35 diseases are increasingly leading to premature deaths <sup>12</sup>. Plant breeders aim to maximize 36 levels of such beneficial compounds, but with little genomic information about rocket species presently available, this is a formidable task. This review will give an overview of research in 37 38 rocket, an outbreeding crop, and how breeders and processors can utilize it to enhance beneficial compounds. 39

#### 40 Rocket species

41 Rocket (also known as arugula, rucola and roquette) is a leafy vegetable crop that has gained 42 substantial popularity across the world, particularly over the last fifteen years <sup>13-16</sup>. Two main 43 species are predominantly farmed as salad crops; these are *Eruca sativa* ('salad' or 'cultivated' 44 rocket; sometimes referred to as Eruca vesicaria subsp. sativa) and Diplotaxis tenuifolia ('wild' 45 rocket). Both species share a peppery taste and aroma that is very distinctive <sup>17</sup>. They have been reported to contain high levels of vitamin C, GSLs, flavonols and phenolics <sup>18-25</sup>. These 46 are all known to have both anti-oxidant and anti-cancer properties, and are also implicated in 47 48 lowering the risk of cardiovascular and cognitive disease. For excellent information on these 49 beneficial effects and their underlying causes, see Drewnowski & Gomez-Carneros <sup>26</sup>, Keum et.al <sup>27</sup>, D'Antuono et.al <sup>28</sup>, Egea-Gilbert et.al <sup>29</sup>, Degl'Innoocenti et.al <sup>30</sup>, Bjorkman et.al <sup>31</sup> and
Jeffery et.al <sup>32</sup>.

#### 52 **Taxonomy and domestication**

A distinction should be made that both *Eruca* and *Diplotaxis* species have overlapping 53 54 characteristics, and that one can be easily mistaken for the other by the untrained eye, and/or before a certain level of maturity has been reached <sup>28</sup>. It is also arguable that *D. tenuifolia* is 55 the least 'wild' of the two species even though the common name is 'wild rocket'. It is featured 56 57 and favored in commercial products and breeding programs, and is likely to be more 58 domesticated than Eruca species as a result. Diplotaxis varieties are generally uniform 59 phenotypically, with *Eruca* varieties being more diverse in this respect <sup>23</sup>. No direct genomic 60 evidence has been presented in the literature to suggest one species is any more or less 61 genetically variable than the other. Variability in GSL data seems to support the hypothesis 62 that *Diplotaxis* species are more 'wild' <sup>33</sup>, though it is not conclusive, as only a relatively small number of cultivars have been tested. This is a point that needs clarification through research 63 64 and extensive breeding, as neither species can be considered fully domesticated <sup>29</sup>. For 65 example, germination rates are variable, reproductive organs are typically small, seedpods 66 shatter and disperse freely (rather than staying on the plant), and physical defenses such as leaf hairs are still present in many commercial varieties <sup>34</sup>. 67

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#### 69 Phytochemicals in *Eruca sativa* and *Diplotaxis tenuifolia*: types and structures

70 Glucosinolates

GSLs are  $\beta$ -thioglucoside *N*-hydrosulphates that are responsible for the sharp and bittertasting flavors found in cruciferous vegetables <sup>35,36</sup>. In combination with the enzyme myrosinase (thioglucoside glucohydrolase, EC 3.2.1.147), GSLs are hydrolyzed to create isothiocyanates, nitriles, thiocyanates, epithionitriles, indoles, oxazolidine-2-thiones,

cyanopithioalkanes, ascorbigens, goitrogens and epithioalkanes <sup>37–49</sup>; see Figure 1. Many of these hydrolysis products have antibacterial, antifungal and insect repellant effects <sup>50–55</sup>. GSLs and ITCs are being increasingly used as 'biofumigants' to suppress soil borne pathogens, nematodes and weeds. Some of the volatile products have the opposite effect of attracting species that can tolerate high GSL concentrations, such as types of ovipositing insect <sup>56,57</sup>.

The conditions under which hydrolysis of GSLs occurs will affect the respective 80 81 proportions of the chemicals produced; pH, iron ions, thiol ions, temperature and hydration play a particularly prominent role in this process *in vivo* <sup>58</sup>. The separation of GSLs in 82 83 specialist 'S-cells' from myrosinase in myrosin cells means that the two components only come into contact upon tissue disruption; for example when damaged via chewing or 84 digestion <sup>59–69</sup>. It is the biological activity of the ITC hydrolysis products in humans that are of 85 86 most interest in rocket <sup>50</sup>. GSLs can be hydrolyzed within the intestinal tract by gut microflora 87 that are known to have specific myrosinase activity <sup>70–73</sup>, but the efficacy of their action is not yet well determined. 88

GSL concentrations can vary and change over time depending on environmental conditions and stress <sup>7</sup>. Other factors affecting GSL profiles include the plant age, organ type, developmental stage, ambient air temperature, level of water stress, photoperiod, agronomic practice, degree of wounding, and geographical origin of the variety/species <sup>74–81</sup>. These can often affect the profiles of *all* phytonutrients contained within tissue, not just GSLs <sup>82</sup>, and they are all factors that plant breeders aim to mitigate through development of genetically advanced and uniform breeding lines.

GSLs and the ITC derivatives have been an integral part of the human diet for millennia
because of the presence of them in the family *Brassicaceae* <sup>64–66,83–89</sup>. GSLs are evolutionarily
recent secondary metabolic products having arisen 10-15 million years ago <sup>90,91</sup>, acting to
prevent pathogen attack and dissuade herbivory. They are known in only a few angiosperm

families of the order *Brassicales*, which includes the *Brassicaceae* <sup>92-100</sup>, and of which *Eruca*and *Diplotaxis* are members.

A study by Pasini et al. <sup>33</sup> of 37 rocket accessions (*Diplotaxis* and *Eruca*) showed that GSL profiles were all very similar, regardless of the species. In total, twelve GSL compounds were found across all accessions; Table 1 illustrates all known GSL compounds identified todate in rocket. These include 4-mercaptobutyl GSL (glucosativin) <sup>21</sup>, 4-methylthiobutyl GSL (glucoerucin) <sup>101</sup>, and 4-methylsulfinylbutyl GSL (glucoraphanin) <sup>28</sup>, which constitute the three most abundant GSLs in rocket.

#### 108 Flavonols

109 Flavonols are diphenylpropanes  $(C_6-C_3-C_6)^{102}$  and are another important group of chemicals found within rocket species. Flavonols in rocket are found with sugar conjugates, and 110 111 typically occur in relatively large quantities <sup>103</sup>. The aglycones found (such as quercetin and 112 kaempferol) are glycosylated and acylated, which in turn affects their biological properties <sup>18</sup>. A study by Martínez-Sánchez et al. <sup>18</sup> identified over 50 different flavonol compounds across 113 114 four different species. Watercress, mizuna and two species of rocket were all found to 115 accumulate very different compounds within their leaves, and in varying quantities. Wild 116 rocket showed high levels of quercetin-3,3',4-triglucosyl (43.5 mg per 100g<sup>-1</sup> fw) and salad 117 rocket had mostly kaempferol-3,4'-diglucosyl (97.8mg per 100g-1 fw). The group also showed 118 a correlation between quercetin derivatives and high antioxidant activity, despite the 119 significant variations seen between species.

120 Studies conducted on rocket tissues have identified significant concentrations of 121 polyglycosylated flavonols. The core aglycones of these are kaempferol, quercetin and 122 isorhamnetin <sup>102</sup>; Table 2 provides an up-to-date list of all flavonol compounds identified in 123 rocket to-date. Martinez-Sanchez et al. <sup>18</sup> showed that *Eruca* species accumulate kaempferol 124 derivatives, whereas *D. tenuifolia* accumulates predominantly quercetin instead, meaning that

the two chemicals could be used as an identification marker between the two species <sup>104</sup>.
Isorhamnetin aglycones are common to both species but typically in much lower
concentrations <sup>33</sup>. The specific aglycones also infer varying degrees of anti-oxidant activity.
For example, quercetin derivatives have a higher activity than kaempferol and isorhamnetin.
The differences in structure (the arrangement of hydroxyl groups and glycosylation) affect
anti-oxidant activity by allowing the molecules to act as hydrogen/electron donors, single
oxygen scavengers, or as reducing agents <sup>105</sup>.

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#### 133 **Phytochemicals and the relation with quality: taste and aroma**

134 It is thought that the presence of glucosativin, glucoerucin and their hydrolysis products 135 within rocket leaves is what gives them a characteristic flavor <sup>44</sup>. Many of the health beneficial 136 GSLs and ITCs are thought to be responsible for strong tastes that some consumers find 137 repellant <sup>106</sup>. It seems that to many people, these compounds contribute very little to a pleasurable eating experience and are actively avoided <sup>83</sup>. Conversely however, some people 138 139 do prefer these strong tastes and aromas, and will actively seek to consume rocket when it is 140 available. Growers in Italy often prefer the subsequent cuts because of the more intense tastes 141 and aromas that are produced <sup>107</sup> and some will even 'sacrifice' the first cut in favour of the 142 subsequent leaf growth. This highlights a divide between consumers that may be indicative of 143 underlying genotype(s) for taste perception and preference.

The breeding process in rocket varieties to-date has effectively made the species 'milder' in taste when compared to plants that grow naturally in the wild. Whether this has been intentional or as a result of selecting for other unrelated traits (such as leaf morphology) is debatable. Some recent commercial varieties have been bred for a 'hotter' taste, such as '*Wildfire*', by Tozer Seeds (Surrey, UK).

149 A study by Pasini et al. <sup>17</sup> demonstrated how breeding for sensory traits could be 150 achieved, by highlighting which glucosinolates contributed to specific taste and aroma 151 elements in rocket. It was found that progoitrin/epiprogoitrin is responsible for bitter taste 152 attributes, despite being only a minor component of the overall GSL profile of rocket (4.3-153 11.4% of total GSL concentration). The perceived pungency of leaves was positively related to the overall GSL content of accessions, and the levels of glucoraphanin negatively contributed 154 155 to the typical 'rocket' flavour. The study also highlighted an important difference between rocket and other *Brassica* sensory studies <sup>108</sup>, in that bitterness was perceived as a favorable 156 157 characteristic according to panelists. The flavonol compound kaempferol-3-(2-sinapoylglucoside)-4'-glucoside also significantly and positively contributed to flavor attributes in 158 159 *Eruca* accessions. This would indicate that GSL compounds are not totally responsible for 160 flavor in rocket. The study itself stopped short of saying how or if the information obtained 161 would be used in breeding programs, but with study into rocket flavor components, milder (and/or stronger) varieties could be bred more efficiently once the responsible compounds 162 163 are properly identified <sup>26</sup>.

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Health promoting properties of glucosinolate-myrosinase products and flavonols of
 rocket

167 Isothiocyanates

168 ITC hydrolysis products have been identified in rocket <sup>45</sup>, such as 4-(methylthio)butyl ITC 169 (erucin) <sup>109,110</sup> which is known to show anti-proliferative activity in human lung carcinoma 170 A549 cells, hepatoma (HepG2) cells, colon cancer cells, prostate cancer cell lines (PC3, BPH-1 171 and LnCap) and leukemia cells <sup>111</sup>. Erucin is a structurally reduced analog of sulforaphane, 172 (which is predominantly found in broccoli) and has shown promising anti-cancer properties 173 *in vitro* (e.g. anti-proliferation of human erytroleukemic K562 cells) <sup>112</sup>. Research into the

174 chemopreventative and anti-genotoxic nature of ITCs has shown promising results <sup>113</sup> (see 175 Figure 2). Other studies involving chemically induced genotoxicity have shown very strong 176 anti-genotoxic effects of *E. sativa* extracts <sup>13</sup> which is in agreement with other *Brassicaceae* 177 studies <sup>114,115</sup>. Identifying specific cultivars of rocket with elevated levels of erucin and 178 glucoraphanin would be an important first-step in developing superior varieties from a 179 human nutrition standpoint.

180 The results of GSL/ITC research prompted an investment in broccoli breeding in the 181 last decade. A similar concerted effort could be made for rocket which contains similar compounds, and which are potentially just as efficacious in humans <sup>116</sup>. Erucin for example, 182 183 has been shown to have very similar, and even superior, biological activity to sulforaphane <sup>117</sup>. 184 One paper has specifically demonstrated that the concentrations of rocket ingested in an average daily diet is significant enough to infer a cancer preventative effect <sup>13</sup>. The 185 186 metabolism of ITCs in humans via the mercapturic acid pathway has been investigated. ITCs 187 are conjugated with glutathione and degraded by N-acetylation, initiating an increase of phase 188 II detoxification enzymes; see Figure 3 for detailed pathway breakdown of erucin <sup>113</sup>.

#### 189 Nitriles

190 Along with ITCs, nitriles are the most abundant bioactive compounds produced by GSL hydrolysis <sup>116</sup>. The hydrolysis of glucoraphanin for example, yields predominantly 191 192 sulforaphane and sulforaphane nitrile. The ratio in which the two are formed depends greatly upon the environmental conditions and the plant cultivar that is used <sup>117</sup>. A low pH medium 193 194 tends towards the formation of nitriles, whereas high pH forms ITCs <sup>118,119</sup>. The presence of 195 thiol and iron ions favors nitriles, and high temperature and hydration produce more ITCs 196 <sup>120,121</sup>. This can have substantial consequences for any potential health benefits that might be 197 inferred from eating rocket <sup>119</sup>. The nitrile form is approximately three orders of magnitude 198 less efficacious than the ITC in inducing quinone reductase (phase II enzyme), and thus infers

a reduced enzymatic and anticarcinogenic response. Nitriles also compete with ITCs in this
induction, and reduce potential positive effects further <sup>38</sup>. As the ratio of these compounds
may depend on plant variety, care must be taken in rocket breeding when selecting plants for
GSL content, as this may not be reflective of the bioactives produced in subsequent hydrolysis
reactions <sup>122</sup>. Other underlying genetic factors may influence which degradation pathway is
taken.

#### 205 Indoles

206 Indoles are the predominant autolysis product of indole glucosinolates such as glucobrassicin, 207 as their ITC counterparts are unstable <sup>85</sup>. Glucobrassicin has been detected as a minor GSL in rocket species <sup>33</sup>, and the predominant indole species produced is indole-3-carbinol. This 208 209 compound is known to be cancer-preventative <sup>125,126</sup>, particularly in reproductive organs *in* 210 vitro and in vivo. A condensation product of indole-3-carbinol, 3,3'-diindolymethane, is also 211 responsible for beneficial physiological effects. Both compounds have been shown to reduce cell proliferation in breast, prostate, cervical and colon cancer cell lines. They also show 212 213 distinct differences from ITCs such as sulforaphane <sup>127</sup>, and inhibition of tumor development 214 in the stomach, breast, uterus, tongue and liver of rodents <sup>128–135</sup>. Experiments in rodents have 215 shown an increase in drug-metabolizing enzymes in the stomach, liver and small intestines of 216 individuals consuming both ITCs and indoles. This is suggestive of enhanced detoxification 217 phase II enzymes (such as quinone reductase, glutathione reductase and glutathione 218 transferase) <sup>134</sup>, and a mechanism by which these phytochemicals infer chemopreventative 219 effects 135,136.

Typically indoles inhibit cell proliferation through cytostatic mechanisms, whereas ITCs induce cytotoxicity within cell lines (at above 12.5µM concentrations), which ultimately leads to increased apoptosis <sup>137,138</sup>. This indicates that both types of compound could act and be effective at different stages of cancer development <sup>11</sup>. Indoles have been shown to induce

programmed cell death in prostate, breast and osteocarcinoma cell lines  $^{139}$  and  $G_1$  cell cycle arrest in breast and prostate cancer cell lines  $^{142,143}$ . It is these cytostatic effects on cell proliferation that has been suggested as the mechanism responsible for the lack of apoptosis effects in indoles  $^{141}$ .

Using information on GSL content in rocket, the ITC and indole effects can be potentially maximized in new varieties, and be of a greater benefit to human health when considered in tandem, rather than separately <sup>127</sup>.

#### 231 Oxazolidine-2-thiones & goitrogens

232 The hydrolysis of  $\beta$ -hydroxy-alkyl GSL compounds (e.g. progoitrin; a minor GSL in rocket) can produce oxazolidine-2-thiones such as goitrin (5-vinyloxazolidine-2-thione) <sup>142-148</sup>. It is these 233 compounds that are largely attributed to the thyroid condition of goiter in mammals <sup>149</sup>, but 234 235 the action of microflora in the gut is thought to mediate the problems associated with high oxazolidine-2-thione intake <sup>150,151</sup>. That being said, oxazolidine-2-thiones interfere with 236 237 thyroxine synthesis <sup>154</sup> and are therefore likely to have an adverse biological effect regardless 238 of gut microflora action or bodily iodine status <sup>3</sup>. A study by Nishie and Daxenbilcher <sup>155</sup> 239 showed that these compounds are not teratogenic or embryotoxic however.

These molecules contribute significantly to the bitter taste of rocket that some people perceive quite strongly <sup>154</sup>. The detection of these compounds may be mediated in a similar genetic fashion as PROP (propylthiouracil), for example <sup>155,156</sup>. By using phytochemical data in rocket breeding programs these oxazolidine-2-thione components could be reduced, potentially improving consumer acceptance (depending on the target consumer) and avoiding any possible adverse health effects associated with over-consumption.

#### 246 Ascorbigens

Ascorbigens are formed via the reaction of indole-3-carbinol and 3,3'-diindolymethane with ascorbic acid in the stomach during myrosinase-catalyzed degradation of indoly-3methyl

glucosinolates <sup>157,158</sup>. In this manner it is thought that ascorbigens have a role in cancermodulation <sup>159</sup> via quinone reductase induction <sup>114</sup>. As has been highlighted previously, this has important implications for breeding for plant varieties with enhanced chemopreventative effects.

#### 253 Epithioalkanes

Epithioalkanes are formed as part of the myrosinase reaction with GSLs at low pH with epithiospecifier protein and ferrous ions. These GSLs typically have a side-chain with a double bond, such as sinigrin <sup>160,161</sup>. It is uncertain whether these compounds produce any significant bioactive effect in humans, but the ratio in which they are produced alongside ITCs, nitriles and indoles may impact on these compounds' efficacy as anti-carcinogens.

#### 259 Flavonols

The antioxidant and anti-inflammatory function of flavonols in the human diet are well known and include protecting the colonic epithelium from free radical damage <sup>164–167</sup>. They can induce the up-regulation of enzymes (such as cytochrome P450), that may lead to a decreased risk of cancer, cardiovascular disease, immune dysfunction, atherosclerosis and chronic inflammation <sup>168,169</sup>.

265

#### 266 Factors affecting phytochemical content

#### 267 Breeding and cultivation

Rocket has been consistently shown to be a good dietary source for flavonols, GSLs and antioxidants. However, there can be large differences between plants of the same germplasm accession due to a combination of genetic and environmental variability. This is probably due to the outbreeding nature of the species <sup>104</sup> and a lack of overall uniformity in varieties. Commercial varieties cannot be considered truly domesticated because of this tendency for outcrossing, and the susceptibility of plants to inbreeding depression (a loss of genetic

variability due to repeated self-pollination or crossing with a closely related individual).
Development of advanced open-pollinating breeding lines (lines that are allowed to crosspollinate freely in a population of selected individuals), or even F<sub>1</sub> hybrids (superior varieties
produced by crossing distinctly different, elite inbred lines), could potentially minimize such
variation.

Throughout the food chain there are many aspects that can have an adverse effect on GSL levels within leaves (Figure 4). These include the cultivar choice, cultivation practice, climatic conditions, photoperiod, sulphur and nitrogen availability, harvest date, time spent in storage, the temperature of wash water, levels of physical damage to leaves, packaging atmosphere and food preparation methods <sup>30-32,170-173</sup>.

#### 284 Harvesting

285 Rocket species have the ability to re-grow their leaves repeatedly after cutting, which allows 286 for several harvests to take place under optimal conditions <sup>107</sup>. In parts of southern Italy, it is not unheard of for up to seven harvests to occur from a single planting. This has obvious cost-287 288 saving benefits for growers, but multiple harvests also induce stress responses in rocket that 289 may be detrimental to the flavor and aesthetics of the crop. Stress drives up the production of 290 secondary metabolites such as GSLs and anthocyanins, which will produce very strong, bitter 291 tastes. There are other detrimental effects of multiple harvests; leaves become progressively 292 smaller and more 'skeletal' in appearance with each cutting, for example. High anthocyanin 293 levels also affect the color of leaves, turning them an undesirable pink, purple or red. Color 294 has been found to be one of the most important characteristics consumers look for in rocket 295 <sup>174</sup>, and so the loss of fresh appearance can ultimately lead to rejection of crops by 296 supermarkets and processors.

#### 297 Industrial and culinary processing

298 There are five main influences that have been identified in affecting GSL levels during 299 processing <sup>94</sup>. These are the action of myrosinase hydrolysis, myrosinase inactivation, the 300 lysis and leaching of GSLs into wash-water, thermal degradation of GSLs, and the loss of 301 ascorbic acid, iron and other enzyme co-factors. Myrosinase inactivation and thermal 302 degradation of GSLs is probably less of an issue in rocket species, as the leaves are not 303 typically cooked. The leaves are not ordinarily frozen, and so freeze-thaw hydrolysis is not 304 likely to be a major factor either. Other factors almost certainly play a significant role in GSL 305 and phytochemical loss in rocket. Verkerk et al. 94 highlighted four key areas that affect GSL 306 levels before reaching the end consumer. These are:

307 1. The variety / cultivar used

308 2. Storage and packaging (post-harvest, post-processing & in shops/supermarkets)

309 3. Industrial processing

310 4. Consumer preparation methods

311 If each of these areas can be mitigated through breeding superior varieties, consumers
312 will receive an end product that is of higher nutritive quality and thus provide increased
313 health benefits.

#### 314 **Post harvest storage**

Studies on both *Diplotaxis* and *Eruca* species have been conducted to determine the effects of post harvest storage conditions on chlorophyll content and respiration rates <sup>15</sup>. Both species of rocket have been found to have high respiration rates <sup>107</sup> leading to rapidly impaired visual quality, such as stem browning, tissue yellowing and general decay <sup>175</sup>. Provided initial GSL loss can be mitigated through breeding, ITC formation has been shown to increase over nitrile formation during the storage period <sup>176</sup>.

Time, temperature, humidity and atmospheric conditions are all optimized for specific
 crops within the logistics chain, but these factors are often only designed to prevent visual

degradation and not phytochemical breakdown <sup>100</sup>. Getting producers, packagers and transporters to change their current practices in order to better preserve the healthpromoting compounds in rocket would be a difficult task. Treatments and storage conditions are often integrated parts of protocols and procedures, and changing these would require significant testing on a commercial scale.

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#### 329 New selection tools for breeders

#### 330 **Phytochemical selection**

331 It should not be forgotten that some GSLs and their breakdown products are thought to be toxic, and even carcinogenic, at high concentrations <sup>128</sup>. Breeders and researchers should be 332 333 mindful that more of a certain compound does not necessarily mean 'better' <sup>177</sup>. Humans seem 334 to be able to tolerate GSLs much better than pigs, rats and rabbits for example; but 335 overconsumption of these compounds may have serious health consequences <sup>64</sup> as high doseeffect relationships are as yet unknown in humans <sup>94</sup>. Few papers in GSL research (regardless 336 337 of species) have acknowledged the potential for plant breeders to utilize HPLC/UPLC/LC-338 MS/GC-MS methods within breeding programs to 'monitor' and select plants for their 339 phytochemical content in this manner. These techniques would provide valuable information 340 on breeding lines relatively rapidly, especially for GSL and flavonol breeding <sup>178</sup>. It is not 341 common practice to select rocket plants based on their phytochemical profile at present, but 342 as interest in these compounds increases it will be necessary for breeders to modify their 343 selection criteria and information sources in order to remain competitive in the salad 344 vegetable market <sup>94</sup>. This has been achieved with 'Beneforte' broccoli (Seminis Vegetable 345 Seeds; subsidiary of Monsanto Company, St. Louis, Missouri, USA; www.beneforte.com) for example. It has also been indicated in hybrid varieties of *Brassica* that ITC/nitrile ratios can be 346 347 selected for <sup>179</sup>.

#### 348 Genetic resources and Marker Assisted Breeding

European initiatives (such as the EU GENRES project 'Leafy vegetables germplasm, stimulating use'; http://documents.plant.wur.nl/cgn/pgr/leafyveg/) have included rocket species within their remit, indicating the rising prominence of the species, and the desire for more work to be conducted on them. The germplasm accessions stored in gene banks are a valuable genetic resource for breeders to take advantage of <sup>180</sup>. The accessions contained within these collections are highly variable and have unique visual and sensory characteristics that could be introgressed into breeding lines relatively easily <sup>181</sup>.

356 Genetic information about rocket within the published literature is very scarce. Some 357 molecular marker techniques such as Random Amplification of Polymorphic DNA (RAPD), 358 Inter-Simple Sequence Repeats (ISSR) and Amplified Fragment Length Polymorphisms 359 (AFLP) have been used to analyze morphological characteristics of *Eruca vesicaria*<sup>29</sup>. ISSR and 360 AFLP are relatively robust for screening variable populations and discriminating between 361 cultivars <sup>180</sup> but RAPDs are notoriously unreliable and suffer from a lack of reproducibility 362 and resolution. Perhaps one of the most underutilized marker types is SRAP (Sequence 363 Related Amplified Polymorphism). The forward and reverse primers are designed to target 364 arbitrary GC and AT rich sequences of the genome respectively, and are therefore more likely to anneal to active genomic regions <sup>182</sup>. This could be of use in understudied crops such as 365 366 rocket, as it provides a simple, repeatable and reliable way of screening large populations.

These techniques are now for the most part however, obsolete in advanced molecular plant breeding, as NGS (Next Generation Sequencing) and SNP (Single Nucleotide Polymorphism)/QTL (Quantitative Trait Loci) analyses are far more specific, reliable and cost-effective. SNPs are the most abundant marker type within genomes, and their high density is ideal for studying specific regions in detail <sup>183</sup>. NGS techniques are now relatively affordable, even for relatively small companies. They are widely available in academic

373 institutions, but many companies are bypassing these in favor of dedicated private commercial services <sup>184</sup> or are developing their own in-house facilities. The inability of some 374 375 research institutions to provide adequate customer service, cost-effectiveness, data storage, 376 and results on time is jeopardizing how much knowledge is in the public domain. Increasingly, 377 both large and small breeding companies are collaborating privately and advancing 378 techniques far beyond those found in academic institutions. Future work by institutes in 379 advanced genomics, sequencing and genotyping is likely to be obsolete in some cases because 380 private research is already finding new innovations, e.g. for data storage and bioinformatics. 381 Because private companies have no obligation to share their knowledge, many of these 382 advances may be unobserved by the mainstream scientific community. Institutes and 383 Universities need to do more to attract business from industry in order to keep up with the 384 pace of private advances in this area.

385 Transcriptome sequences are now (generally) adequate for breeders to use and make huge advances in only a few years. Linkage mapping and QTL analyses can be conducted on 386 387 desktop computers, making integration into breeding companies relatively straightforward 388 from an IT point of view, even if the actual sequencing and genotyping are outsourced. Again, 389 this may typically be to private companies providing a dedicated service. The availability of 390 software licenses and advanced training courses from private companies also means plant 391 breeders do not necessarily need the expertise found in Universities and research institutes in 392 order to attain their goals.

393

#### 394 Summary

Of all the research papers concerning rocket species and their phytochemistry, none have
directly addressed how information could be used within a working breeding population.
Often it is explained or postulated purely as theory rather than actual practice, or only given a

398 cursory mention. Only very rarely is a plant breeding program reflective of theory, due to the 399 large number of environmental factors affecting plant growth, development and reproduction. 400 The progressive selection of rocket plants through conventional/molecular breeding would 401 be a valuable tool for the research community as well as providing an excellent incentive for 402 breeding companies to fund research. The actual monitoring and quantification of 403 GSL/flavonol levels through successive generations (i.e. not just one as has been the case with 404 most studies) would not only validate the heritability of such traits in rocket, but would also 405 provide a 'roadmap' for how other minor crops might be developed for commercial use.

406 Attention must be paid to the phytochemical content of varieties within breeding 407 populations of rocket. By focusing solely on morphological traits, important phytochemical 408 genotypes may be inadvertently lost from populations; this could be said of all *Brassicaceae* 409 species, not just rocket. The balance of glucosinolate-myrosinase degradation products does 410 seem to have a genetic component to it and so could be selected for also. Utilising genetic 411 resources, the falling costs of sequencing and bioinformatics can produce nutritively superior 412 varieties of rocket in the near future. Plant breeding typically takes longer than the average 413 research project allows for, even with the use of advanced genomic selection methods. This is 414 a situation that could be remedied by long-term industrial collaboration and sponsorship by 415 plant breeding firms.

## 416 References417

418 (1) Chaudhary, A.; Rampal, G.; Sharma, U.; Thind, T. S.; Singh, B.; Vig, A. P.; Arora, S. *Med.*419 *Chem. Drug Discov.* 2012, *2*, 30–37.

420 (2) Gross, H. B.; Dalebout, T.; Grubb, C. D.; Abel, S. *Plant Sci.* **2000**, *159*, 265–272.

421 (3) Jongen, W. M. F. *Proc. Nutr. Soc.* **1996**, *55*, 433–446.

- 422 (4) Vinson, J. A.; Dabbagh, Y. A.; Serry, M. M.; Jang, J. H. *J. Agric. Food Chem.* **1995**, *43*, 2800–
  423 2802.
- 424 (5) Clarke, J. D.; Dashwood, R. H.; Ho, E. *Cancer Lett.* **2008**, *269*, 291–304.

- 425 (6) Hayes, J. D.; Kelleher, M. O.; Eggleston, I. M. Eur. J. Nutr. 2008, 47, 73–88.
- 426 (7) Herr, I.; Buechler, M. W. *Cancer Treat. Rev.* **2010**, *36*, 377–383.
- 427 (8) Melchini, A.; Traka, M. H. *Toxins (Basel).* **2010**, *2*, 593–612.
- 428 (9) Zhang, Y. S. Mutat. Res. Mol. Mech. Mutagen. 2004, 555, 173–190.
- 429 (10) Yang, Y. M.; Conaway, C. C.; Chiao, J. W.; Wang, C. X.; Amin, S.; Whysner, J.; Dai, W.;
  430 Reinhardt, J.; Chung, F. L. *Cancer Res.* 2002, *62*, 2–7.
- 431 (11) Pappa, G.; Lichtenberg, M.; Iori, R.; Barillari, J.; Bartsch, H.; Gerhauser, C. *Mutat. Res. Mol.*432 *Mech. Mutagen.* 2006, 599, 76–87.
- 433 (12) Casagrande, S. S.; Wang, Y.; Anderson, C.; Gary, T. L. *Am. J. Prev. Med.* **2007**, *32*, 257–263.
- 434 (13) Lamy, E.; Schroder, J.; Paulus, S.; Brenk, P.; Stahl, T.; Mersch-Sundermann, V. *Food Chem.* 435 *Toxicol.* 2008, 46, 2415–2421.
- 436 (14) D'Antuono, L. F.; Elementi, S.; Neri, R. J. Sci. Food Agric. 2009, 89, 713–722.
- 437 (15) Koukounaras, A.; Siomos, A. S.; Sfakiotakis, E. *Postharvest Biol. Technol.* 2007, 46, 167–
  438 173.
- 439 (16) Hall, M. K. D.; Jobling, J. J.; Rogers, G. S. Veg. Crop. Res. Bull. 2012, 76, 21–41.
- 440 (17) Pasini, F.; Verardo, V.; Cerretani, L.; Caboni, M. F.; D'Antuono, L. F. *J. Sci. Food Agric.*441 **2011**, *91*, 2858–2864.
- 442 (18) Martinez-Sanchez, A.; Gil-Izquierdo, A.; Gil, M. I.; Ferreres, F. *J. Agric. Food Chem.* 2008, 56, 2330–2340.
- 444 (19) Kim, S. J.; Ishii, G. J. Sci. Food Agric. 2007, 87, 966–973.
- 445 (20) Bennett, R. N.; Carvalho, R.; Mellon, F. A.; Eagles, J.; Rosa, E. A. S. *J. Agric. Food Chem.*446 2007, 55, 67–74.
- 447 (21) Bennett, R. N.; Mellon, F. A.; Botting, N. P.; Eagles, J.; Rosa, E. A. S.; Williamson, G.
  448 *Phytochemistry* 2002, *61*, 25–30.
- 449 (22) Cataldi, T. R. I.; Rubino, A.; Lelario, F.; Bufo, S. A. *Rapid Commun. Mass Spectrom.* 2007, 21, 2374–2388.
- 451 (23) Bennett, R. N.; Rosa, E. A. S.; Mellon, F. A.; Kroon, P. A. J. Agric. Food Chem. 2006, 54,
  452 4005–4015.
- 453 (24) Chun Arasu, M.V., Lim, Y-P., Kim, S-J., J.-H. *Hortic. Environ. Biotechnol.* 2013, 54, 206–
  454 213.

- 455 (25) Martinez-Sanchez, A.; Llorach, R.; Gil, M. I.; Ferreres, F. *J. Agric. Food Chem.* 2007, *55*,
  456 1356–1363.
- 457 (26) Drewnowski, A.; Gomez-Carneros, C. Am. J. Clin. Nutr. **2000**, 72, 1424–1435.
- 458 (27) Keum, Y. S.; Jeong, W. S.; Kong, A. N. T. *Mutat. Res. Mol. Mech. Mutagen.* 2004, 555, 191–
   459 202.
- 460 (28) D'Antuono, L. F.; Elementi, S.; Neri, R. *Phytochemistry* **2008**, 69, 187–199.
- 461 (29) Egea-Gilabert, C.; Fernandez, J. A.; Migliaro, D.; Martinez-Sanchez, J. J.; Vicente, M. J. *Sci.* 462 *Hortic. (Amsterdam).* 2009, *121*, 260–266.
- 463 (30) Degl'Innoocenti, E.; Pardossi, A.; Tattini, M.; Guidi, L. *J. Food Biochem.* 2008, *32*, 642–
  464 653.
- 465 (31) Bjorkman, M.; Klingen, I.; Birch, A. N. E.; Bones, A. M.; Bruce, T. J. A.; Johansen, T. J.;
  466 Meadow, R.; Molmann, J.; Seljasen, R.; Smart, L. E.; Stewart, D. *Phytochemistry* 2011, 72,
  467 538–556.
- 468 (32) Jeffery, E. H.; Brown, A. F.; Kurilich, A. C.; Keck, A. S.; Matusheski, N.; Klein, B. P.; Juvik, J.
  469 A. *J. Food Compos. Anal.* **2003**, *16*, 323–330.
- 470 (33) Pasini, F.; Verardo, V.; Caboni, M. F.; D'Antuono, L. F. *Food Chem.* **2012**, *133*, 1025–1033.
- 471 (34) Gepts, P. What is a crop?: The Domestication Syndrome.
  472 http://www.plantsciences.ucdavis.edu/gepts/pb143/LEC16/Pb143l16.htm
- 473 (35) Rungapamestry, V.; Duncan, A. J.; Fuller, Z.; Ratcliffe, B. *Proc. Nutr. Soc.* **2007**, *66*, 69–81.
- 474 (36) Velasco, P.; Cartea, M. E.; Gonzalez, C.; Vilar, M.; Ordas, A. *J. Agric. Food Chem.* 2007, *55*,
  475 955–962.
- 476 (37) Hecht, S. S. J. Nutr. **1999**, *129*, 768S–774S.
- 477 (38) Matusheski, N. V; Jeffery, E. H. J. Agric. Food Chem. **2001**, 49, 5743–5749.
- 478 (39) Rangkadilok, N.; Nicolas, M. E.; Bennett, R. N.; Premier, R. R.; Eagling, D. R.; Taylor, P. W.
  479 J. *Sci. Hortic. (Amsterdam).* 2002, *96*, 27–41.
- 480 (40) Yuan, G. F.; Sun, B.; Yuan, J.; Wang, Q. M. J. Zhejiang Univ. B **2009**, *10*, 580–588.
- 481 (41) Jia, C. G.; Xu, C. J.; Wei, J.; Yuan, J.; Yuan, G. F.; Wang, B. L.; Wang, Q. M. Food Chem. 2009, 114, 28–37.
- 483 (42) McNaughton, S. A.; Marks, G. C. Br. J. Nutr. 2003, 90, 687–697.
- 484 (43) Zhang, Y. S.; Talalay, P.; Cho, C. G.; Posner, G. H. *Proc. Natl. Acad. Sci. U. S. A.* **1992**, *89*,
  485 2399–2403.

- 486 (44) Bones, A. M.; Rossiter, J. T. *Phytochemistry* **2006**, *67*, 1053–1067.
- 487 (45) Jirovetz, L.; Smith, D.; Buchbauer, G. J. Agric. Food Chem. 2002, 50, 4643–4646.
- 488 (46) Grubb, C. D.; Abel, S. *Trends Plant Sci.* **2006**, *11*, 89–100.
- 489 (47) Yan, X. F.; Chen, S. X. *Planta* **2007**, *226*, 1343–1352.
- 490 (48) Baik, H. Y.; Juvik, J.; Jeffery, E. H.; Wallig, M. A.; Kushad, M.; Klein, B. P. *J. Food Sci.* 2003, 68, 1043–1050.
- 492 (49) Taiz, L.; Zeiger, E. *Plant physiology*; 4th ed.; Sinauer: Sunderland, Mass, 2006; p. xxvi,
  493 764 p.
- 494 (50) Halkier, B. A.; Gershenzon, J. In *Annual Review of Plant Biology*; 2006; Vol. 57, pp. 303–
   495 333.
- 496 (51) Mithen, R.; Campos, H. *Entomol. Exp. Appl.* **1996**, *80*, 202–205.
- 497 (52) Newman, R. M.; Hanscom, Z.; Kerfoot, W. C. *Oecologia* **1992**, *92*, 1–7.
- 498 (53) Ostrofsky, M. L.; Zettler, E. R. J. Ecol. 1986, 74, 279–287.
- 499 (54) Jeffries, M. Freshw. Biol. 1990, 23, 265–269.
- 500 (55) Newman, R. M.; Kerfoot, W. C.; Hanscom, Z. J. Chem. Ecol. **1990**, *16*, 245–259.
- 501 (56) Brown, P. D.; Morra, M. J. J. Agric. Food Chem. **1995**, 43, 3070–3074.
- 502 (57) Vaughn, S. F.; Isbell, T. A.; Weisleder, D.; Berhow, M. A. J. Chem. Ecol. 2005, 31, 167–177.
- 503 (58) Foo, H. L.; Gronning, L. M.; Goodenough, L.; Bones, A. M.; Danielsen, B. E.; Whiting, D. A.;
   504 Rossiter, J. T. *FEBS Lett.* 2000, *468*, 243–246.
- 505 (59) Song, L.; Thornalley, P. J. Food Chem. Toxicol. 2007, 45, 216–224.
- 506 (60) Verkerk, R.; Dekker, M.; Jongen, W. M. F. J. Sci. Food Agric. 2001, 81, 953–958.
- 507 (61) Andreasson, E.; Jorgensen, L. B.; Hoglund, A. S.; Rask, L.; Meijer, J. *Plant Physiol.* 2001, 127, 1750–1763.
- 509 (62) Husebye, H.; Chadchawan, S.; Winge, P.; Thangstad, O. P.; Bones, A. M. *Plant Physiol.* 510 2002, *128*, 1180–1188.
- 511 (63) Kliebenstein, D. J.; Kroymann, J.; Mitchell-Olds, T. *Curr. Opin. Plant Biol.* 2005, *8*, 264–
   512 271.
- 513 (64) Tripathi, M. K.; Mishra, A. S. Anim. Feed Sci. Technol. 2007, 132, 1–27.
- 514 (65) Talalay, P.; Fahey, J. W. J. Nutr. **2001**, *131*, 3027S–3033S.

- 515 (66) Chen, S.; Andreasson, E. *Plant Physiol. Biochem.* **2001**, *39*, 743–758.
- 516 (67) Hoglund, A. S.; Lenman, M.; Falk, A.; Rask, L. *Plant Physiol.* **1991**, *95*, 213–221.
- 517 (68) Getahun, S. M.; Chung, F. L. *Cancer Epidemiol. Biomarkers Prev.* **1999**, *8*, 447–451.
- 518 (69) Fenwick, G. R.; Heaney, R. K. *Food Chem.* **1983**, *11*, 249–271.
- 519 (70) Rabot, S.; Nugonbaudon, L.; Raibaud, P.; Szylit, O. *Br. J. Nutr.* **1993**, *70*, 323–331.
- 520 (71) Shapiro, T. A.; Fahey, J. W.; Wade, K. L.; Stephenson, K. K.; Talalay, P. *Cancer Epidemiol.*521 *Biomarkers Prev.* 1998, 7, 1091–1100.
- 522 (72) Fahey, J. W.; Zhang, Y. S.; Talalay, P. *Proc. Natl. Acad. Sci. U. S. A.* **1997**, *94*, 10367–10372.
- 523 (73) Heaney, R. K.; Fenwick, G. R. J. Sci. Food Agric. **1980**, 31, 593–599.
- 524 (74) Kushad, M. M.; Brown, A. F.; Kurilich, A. C.; Juvik, J. A.; Klein, B. P.; Wallig, M. A.; Jeffery, E.
  525 H. *J. Agric. Food Chem.* **1999**, *47*, 1541–1548.
- 526 (75) Rosa, E.; Heaney, R. Anim. Feed Sci. Technol. 1996, 57, 111–127.
- 527 (76) Rangkadilok, N.; Nicolas, M. E.; Bennett, R. N.; Premier, R. R.; Eagling, D. R.; Taylor, P. W.
  528 J. *Sci. Hortic. (Amsterdam).* 2002, 96, 11–26.
- 529 (77) Agerbirk, N.; Olsen, C. E.; Nielsen, J. K. *Phytochemistry* **2001**, *58*, 91–100.
- 530 (78) Coogan, R. C.; Wills, R. B. H.; Nguyen, V. Q. Food Chem. **2001**, 72, 1–3.
- 531 (79) Ahuja, I.; de Vos, R. C. H.; Bones, A. M.; Hall, R. D. *Trends Plant Sci.* **2010**, *15*, 664–674.
- 532 (80) Hasegawa, T.; Yamada, K.; Kosemura, S.; Yamamura, S.; Hasegawa, K. *Phytochemistry* 533 2000, 54, 275–279.
- 534 (81) Bartlet, E.; Kiddle, G.; Williams, I.; Wallsgrove, R. *Entomol. Exp. Appl.* **1999**, *91*, 163–167.
- 535 (82) Jin, J.; Koroleva, O. A.; Gibson, T.; Swanston, J.; Magan, J.; Zhang, Y.; Rowland, I. R.;
  536 Wagstaff, C. *J. Agric. Food Chem.* **2009**, *57*, 5227–5234.
- 537 (83) Holst, B.; Williamson, G. Nat. Prod. Rep. 2004, 21, 425–447.
- 538 (84) Stoewsand, G. S. *Food Chem. Toxicol.* **1995**, *33*, 537–543.
- 539 (85) Fahey, J. W.; Zalcmann, A. T.; Talalay, P. *Phytochemistry* **2001**, *56*, 5–51.
- 540 (86) Cartea, M. E.; Francisco, M.; Soengas, P.; Velasco, P. *Molecules* **2011**, *16*, 251–280.
- 541 (87) Zhang, Y. S.; Talalay, P. *Cancer Res.* **1994**, *54*, S1976–S1981.

- 542 (88) Rose, P.; Won, Y. K.; Ong, C. N.; Whiteman, M. *Nitric Oxide-Biology Chem.* 2005, *12*, 237–
  543 243.
- 544 (89) Conaway, C. C.; Yang, Y. M.; Chung, F. L. *Curr. Drug Metab.* **2002**, *3*, 233–255.
- 545 (90) Windsor, A. J.; Reichelt, M.; Figuth, A.; Svatos, A.; Kroymann, J.; Kliebenstein, D. J.;
  546 Gershenzon, J.; Mitchell-Olds, T. *Phytochemistry* 2005, 66, 1321–1333.
- 547 (91) Wheat, C. W.; Vogel, H.; Wittstock, U.; Braby, M. F.; Underwood, D.; Mitchell-Olds, T. *Proc.* 548 *Natl. Acad. Sci. U. S. A.* 2007, *104*, 20427–20431.
- 549 (92) Rodman, J. E.; Karol, K. G.; Price, R. A.; Sytsma, K. J. Syst. Bot. 1996, 21, 289–307.
- 550 (93) Wittstock, U.; Halkier, B. A. *Trends Plant Sci.* **2002**, *7*, 263–270.
- (94) Verkerk, R.; Schreiner, M.; Krumbein, A.; Ciska, E.; Holst, B.; Rowland, I.; De Schrijver, R.;
  Hansen, M.; Gerhauser, C.; Mithen, R.; Dekker, M. *Mol. Nutr. Food Res.* 2009, *53*, S219–
  S265.
- 554 (95) Rosa, E. A. S. *Phytochemistry* **1997**, *44*, 1415–1419.
- 555 (96) Clarke, D. B. Anal. Methods **2010**, *2*, 310–325.
- 556 (97) Mithen, R. F.; Dekker, M.; Verkerk, R.; Rabot, S.; Johnson, I. T. *J. Sci. Food Agric.* 2000, *80*,
   557 967–984.
- 558 (98) Charron, C. S.; Saxton, A. M.; Sams, C. E. J. Sci. Food Agric. 2005, 85, 671–681.
- 559 (99) Schreiner, M. Eur. J. Nutr. 2005, 44, 85–94.
- (100) Schouten, R. E.; Zhang, X. B.; Verkerk, R.; Verschoor, J. A.; Otma, E. C.; Tijskens, L. M. M.;
  van Kooten, O. *Postharvest Biol. Technol.* 2009, *53*, 1–10.
- 562 (101) Graser, G.; Schneider, B.; Oldham, N. J.; Gershenzon, J. Arch. Biochem. Biophys. 2000, 378,
   563 411–419.
- 564 (102) Arabbi, P. R.; Genovese, M. I.; Lajolo, F. M. J. Agric. Food Chem. **2004**, *52*, 1124–1131.
- 565 (103) Podsedek, A. Lwt-Food Sci. Technol. 2007, 40, 1–11.
- 566 (104) Cartea, M. E.; Velasco, P.; Obregon, S.; Padilla, G.; de Haro, A. *Phytochemistry* 2008, 69,
   567 403–410.
- 568 (105) Salah, N.; Miller, N. J.; Paganga, G.; Tijburg, L.; Bolwell, G. P.; Riceevans, C. *Arch. Biochem.* 569 *Biophys.* 1995, *322*, 339–346.
- 570 (106) Hansen, M.; Laustsen, A. M.; Olsen, C. E.; Poll, L.; Sorensen, H. J. Food Qual. 1997, 20,
   571 441–459.

- 572 (107) Martinez-Sanchez, A.; Allende, A.; Cortes-Galera, Y.; Gil, M. I. *Postharvest Biol. Technol.* 573 2008, 47, 382–388.
- 574 (108) Schonhof, I.; Krumbein, A.; Bruckner, B. *Nahrung-Food* **2004**, *48*, 25–33.
- 575 (109) Iori, R.; Bernardi, R.; Gueyrard, D.; Rollin, P.; Polmieri, S. *Bioorg. Med. Chem. Lett.* 1999,
   576 9, 1047–1048.
- 577 (110) Cerny, M. S.; Taube, E.; Battaglia, R. J. Agric. Food Chem. **1996**, 44, 3835–3839.
- 578 (111) Melchini, A.; Costa, C.; Traka, M.; Miceli, N.; Mithen, R.; De Pasquale, R.; Trovato, A. *Food* 579 *Chem. Toxicol.* 2009, 47, 1430–1436.
- 580 (112) Leoni, O.; Iori, R.; Palmieri, S.; Esposito, E.; Menegatti, E.; Cortesi, R.; Nastruzzi, C. *Bioorg.* 581 *Med. Chem.* **1997**, *5*, 1799–1806.
- 582 (113) Wu, X.; Zhou, Q. H.; Xu, K. *Acta Pharmacol. Sin.* **2009**, *30*, 501–512.
- 583 (114) Zhu, C. Y.; Loft, S. *Food Chem. Toxicol.* **2003**, *41*, 455–462.
- 584 (115) Kassie, F.; Rabot, S.; Uhl, M.; Huber, W.; Qin, H. M.; Helma, C.; Schulte-Hermann, R.;
   585 Knasmuller, S. *Carcinogenesis* 2002, *23*, 1155–1161.
- 586 (116) Alqasoumi, S.; Ai-Sohaibani, M.; Ai-Howiriny, T.; Al-Yahya, M.; Rafatullah, S. *World J.* 587 *Gastroenterol.* 2009, *15*, 1958–1965.
- 588 (117) Hanlon, N.; Coldham, N.; Sauer, M. J.; Ioannides, C. *Chem. Biol. Interact.* 2009, 177, 115–
   589 120.
- 590 (118) Cole, R. A. *Phytochemistry* **1976**, *15*, 759–762.
- (119) Matusheski, N. V; Wallig, M. A.; Juvik, J. A.; Klein, B. P.; Kushad, M. M.; Jeffery, E. H. J.
   *Agric. Food Chem.* 2001, 49, 1867–1872.
- 593 (120) Uda, Y.; Kurata, T.; Arakawa, N. Agric. Biol. Chem. **1986**, 50, 2735–2740.
- 594 (121) Macleod, A. J.; Rossiter, J. T. *Phytochemistry* **1986**, *25*, 1047–1051.
- 595 (122) Tookey, H. L.; Wolff, I. A. Can. J. Biochem. **1970**, 48, 1024–&.
- 596 (123) Uda, Y.; Kurata, T.; Arakawa, N. Agric. Biol. Chem. **1986**, 50, 2741–2746.
- 597 (124) Gil, V.; Macleod, A. J. *Phytochemistry* **1980**, *19*, 227–231.
- (125) Cashman, J. R.; Xiong, Y.; Lin, J.; Verhagen, H.; van Poppel, G.; van Bladeren, P. J.; Larsen Su, S.; Williams, D. E. *Biochem. Pharmacol.* **1999**, *58*, 1047–1055.
- 600 (126) Graham, S. *Cancer Res.* **1983**, *43*, 2409–2413.
- 601 (127) Bonnesen, C.; Eggleston, I. M.; Hayes, J. D. *Cancer Res.* **2001**, *61*, 6120–6130.

- 602 (128) Kim, D. J.; Han, B. S.; Ahn, B.; Hasegawa, R.; Shirai, T.; Ito, N.; Tsuda, H. *Carcinogenesis* 603 1997, *18*, 377–381.
- 604 (129) Wattenberg, L. W.; Loub, W. D. *Cancer Res.* **1978**, *38*, 1410–1413.
- 605 (130) Bresnick, E.; Birt, D. F.; Wolterman, K.; Wheeler, M.; Markin, R. S. *Carcinogenesis* 1990,
   606 *11*, 1159–1163.
- 607 (131) Bradlow, H. L.; Michnovicz, J. J.; Telang, N. T.; Osborne, M. P. *Carcinogenesis* 1991, *12*,
   608 1571–1574.
- 609 (132) Kojima, T.; Tanaka, T.; Mori, H. *Cancer Res.* **1994**, *54*, 1446–1449.
- 610 (133) Tanaka, T.; Kojima, T.; Morishita, Y.; Mori, H. Japanese J. Cancer Res. 1992, 83, 835–842.
- 611 (134) Tanaka, T.; Mori, Y.; Morishita, Y.; Hara, A.; Ohno, T.; Kojima, T.; Mori, H. *Carcinogenesis*612 **1990**, *11*, 1403–1406.
- 613 (135) Kim, D. J.; Lee, K. K.; Han, B. S.; Ahn, B.; Bae, J. H.; Jang, J. J. *Japanese J. Cancer Res.* 1994,
  614 *85*, 578–583.
- 615 (136) Staack, R.; Kingston, S.; Wallig, M. A.; Jeffery, E. H. *Toxicol. Appl. Pharmacol.* 1998, 149,
   616 17–23.
- (137) Verhoeven, D. T. H.; Verhagen, H.; Goldbohm, R. A.; vandenBrandt, P. A.; vanPoppel, G. *Chem. Biol. Interact.* **1997**, *103*, 79–129.
- 619 (138) Wattenberg, L. W. *Cancer Res.* **1985**, *45*, 1–8.
- 620 (139) Kuang, Y. F.; Chen, Y. H. Food Chem. Toxicol. 2004, 42, 1711–1718.
- 621 (140) Gamet-Payrastre, L.; Lumeau, S.; Gasc, N.; Cassar, G.; Rollin, P.; Tulliez, J. *Anticancer*.
   622 *Drugs* 1998, 9, 141–148.
- 623 (141) Ge, X. K.; Fares, F. A.; Yannai, S. Anticancer Res. **1999**, *19*, 3199–3203.
- 624 (142) Cover, C. M.; Hsieh, S. J.; Tran, S. H.; Hallden, G.; Kim, G. S.; Bjeldanes, L. F.; Firestone, G. L.
  625 *J. Biol. Chem.* **1998**, *273*, 3838–3847.
- 626 (143) Sarkar, F. H.; Li, Y. W. J. Nutr. **2004**, *134*, 3493S–3498S.
- 627 (144) Lijang, S.; Iori, R.; Thornalley, P. J. J. Sci. Food Agric. 2006, 86, 1271–1280.
- 628 (145) Zhao, D. Y.; Tang, J.; Ding, X. L. *Lwt-Food Sci. Technol.* **2007**, *40*, 439–447.
- 629 (146) Wink, M. Biochemistry of Plant Secondary Metabolites; Wiley-Blackwell, 2010; Vol. 40.
- 630 (147) Greer, M. A. Arch. Biochem. Biophys. **1962**, 99, 369–371.
- 631 (148) Grubb, C. D.; Gross, H. B.; Chen, D. L.; Abel, S. *Plant Sci.* **2002**, *162*, 143–152.

- 632 (149) Ghawi, S. K.; Methven, L.; Niranjan, K. *Food Chem.* **2013**, *138*, 1734–1741.
- 633 (150) Higdon, J. V; Delage, B.; Williams, D. E.; Dashwood, R. H. *Pharmacol. Res.* 2007, *55*, 224–
  634 236.
- (151) Mcdanell, R.; Mclean, A. E. M.; Hanley, A. B.; Heaney, R. K.; Fenwick, G. R. *Food Chem. Toxicol.* **1988**, *26*, 59–70.
- 637 (152) Lanzani, A.; Piana, G.; Piva, G.; Cardillo, M.; Rastelli, A.; Jacini, G. *J. Am. Oil Chem. Soc.*638 **1974**, *51*, 517–518.
- 639 (153) Mawson, R.; Heaney, R. K.; Zdunczyk, Z.; Kozlowska, H. *Food / Nahrung* 1993, *37*, 336–
  640 344.
- 641 (154) Dewick, P. M. Medicinal natural products a biosynthetic approach, 2009, 1 online
   642 resource (x, 539 p.).
- 643 (155) Nishie, K.; Daxenbichler, M. E. *Food Cosmet. Toxicol.* **1980**, *18*, 159–172.
- 644 (156) Fenwick, G. R.; Griffiths, N. M. Zeitschrift fur Leb. und -forsch. **1981**, 172, 90–92.
- 645 (157) Fenwick, G. R.; Griffiths, N. M.; Heaney, R. K. J. Sci. Food Agric. **1983**, 34, 73–80.
- 646 (158) Lewis, J.; Fenwick, G. R. *Food Chem.* **1987**, *25*, 259–268.
- 647 (159) Buskov, S.; Hansen, L. B.; Olsen, C. E.; Sørensen, J. C.; Sørensen, H.; Sørensen, S. *J. Agric.* 648 *Food Chem.* 2000, *48*, 2693–2701.
- 649 (160) Hrncirik, K.; Valusek, J.; Velisek, J. *Eur. Food Res. Technol.* **2001**, *212*, 576–581.
- (161) Preobrazhenskaya, M. N.; Bukhman, V. M.; Korolev, A. M.; Efimov, S. A. *Pharmacol. Ther.* **1993**, *60*, 301–313.
- (162) Smith, T. K.; Lund, E. K.; Clarke, R. G.; Bennett, R. N.; Johnson, I. T. *J. Agric. Food Chem.* 2005, *53*, 3895–3901.
- (163) Lambrix, V.; Reichelt, M.; Mitchell-Olds, T.; Kliebenstein, D. J.; Gershenzon, J. *Plant Cell* **2001**, *13*, 2793–2807.
- 656 (164) Hollman, P. C. H.; Katan, M. B. *Biomed. Pharmacother.* **1997**, *51*, 305–310.
- 657 (165) Hollman, P. C. H.; Katan, M. B. Food Chem. Toxicol. 1999, 37, 937–942.
- 658 (166) Harborne, J. B.; Williams, C. A. *Phytochemistry* **2000**, *55*, 481–504.
- (167) Olsson, L. C.; Veit, M.; Weissenbock, G.; Bornman, J. F. *Phytochemistry* **1998**, 49, 1021–
  1028.
- (168) Manach, C.; Scalbert, A.; Morand, C.; Remesy, C.; Jimenez, L. *Am. J. Clin. Nutr.* 2004, *79*,
   727–747.

- (169) Kroon, P. A.; Clifford, M. N.; Crozier, A.; Day, A. J.; Donovan, J. L.; Manach, C.; Williamson,
   G. *Am. J. Clin. Nutr.* 2004, *80*, 15–21.
- 665 (170) Engelen-Eigles, G.; Holden, G.; Cohen, J. D.; Gardner, G. *J. Agric. Food Chem.* 2006, *54*,
   666 328–334.
- 667 (171) Palaniswamy, U.; McAvoy, R.; Bible, B. *Hortscience* **1997**, *32*, 222–223.
- 668 (172) Aires, A.; Rosa, E.; Carvalho, R. J. Sci. Food Agric. 2006, 86, 1512–1516.
- 669 (173) Palaniswamy, U.; McAvoy, R.; Bible, B.; Singha, S.; Hill, D. *Phytochem. Heal.* **1995**, *15*,
  670 280–283.
- 671 (174) Koukounaras, A.; Siomos, A. S.; Sfakiotakis, E. J. Food Qual. **2010**, *33*, 768–779.
- (175) Koukounaras, A.; Siomos, A. S.; Sfakiotakis, E. *Postharvest Biol. Technol.* 2006, 41, 109–
   111.
- 674 (176) Howard, L. A.; Jeffery, E. H.; Wallig, M. A.; Klein, B. P. J. Food Sci. 1997, 62, 1098-+.
- (177) Kassie, F.; Parzefall, W.; Musk, S.; Johnson, I.; Lamprecht, G.; Sontag, G.; Knasmuller, S.
   *Chem. Biol. Interact.* **1996**, *102*, 1–16.
- 677 (178) Rochfort, S. J.; Trenerry, V. C.; Imsic, M.; Panozzo, J.; Jones, R. *Phytochemistry* 2008, 69,
   678 1671–1679.
- 679 (179) Faulkner, K.; Mithen, R.; Williamson, G. *Carcinogenesis* **1998**, *19*, 605–609.
- 680 (180) Xu, Y. Molecular Plant Breeding; CABI, 2010.
- (181) Bozokalfa, M. K.; Yagmur, B.; Ilbi, H.; Esiyok, D.; Kavak, S. *Crop Breed. Appl. Biotechnol.* 2009, 9, 372–381.
- 683 (182) Li, G.; Quiros, C. F. *Theor. Appl. Genet.* **2001**, *103*, 455–461.
- 684 (183) Baird, N. A.; Etter, P. D.; Atwood, T. S.; Currey, M. C.; Shiver, A. L.; Lewis, Z. A.; Selker, E.
   685 U.; Cresko, W. A.; Johnson, E. A. *PLoS One* **2008**, *3*.
- 686 (184) Glenn, T. C. *Mol. Ecol. Resour.* **2011**, *11*, 759–769.
- 687 (185) Hall, C.; McCallum, D.; Prescott, A.; Mithen, R. *Theor. Appl. Genet.* **2001**, *102*, 369–374.
- 688 (186) Lelario, F.; Bianco, G.; Bufo, S. A.; Cataldi, T. R. I. *Phytochemistry* **2012**, *73*, 74–83.
- 689 (187) Botting, C. H.; Davidson, N. E.; Griffiths, D. W.; Bennett, R. N.; Botting, N. P. J. Agric. Food
   690 Chem. 2002, 50, 983–988.
- 691 (188) Kim, S. J.; Ishii, G. Soil Sci. Plant Nutr. **2006**, *52*, 394–400.

- (189) Villatoro-Pulido, M.; Priego-Capote, F.; Alvarez-Snachez, B.; Saha, S.; Philo, M.; Obregon Cano, S.; De Haro-Bailon, A.; Font, R.; Del Rio-Celestino, M. J. Sci. Food Agric. 2013.
- 694

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#### **Figure captions**

**Figure 1:** – The glucosinolate-myrosinase reaction and some of the subsequent compounds produced under different conditions, such as pH and the influence of epithiospecifier proteins (ESP) (Adapted from Zhang <sup>9</sup> and Hall et al. <sup>185</sup>).

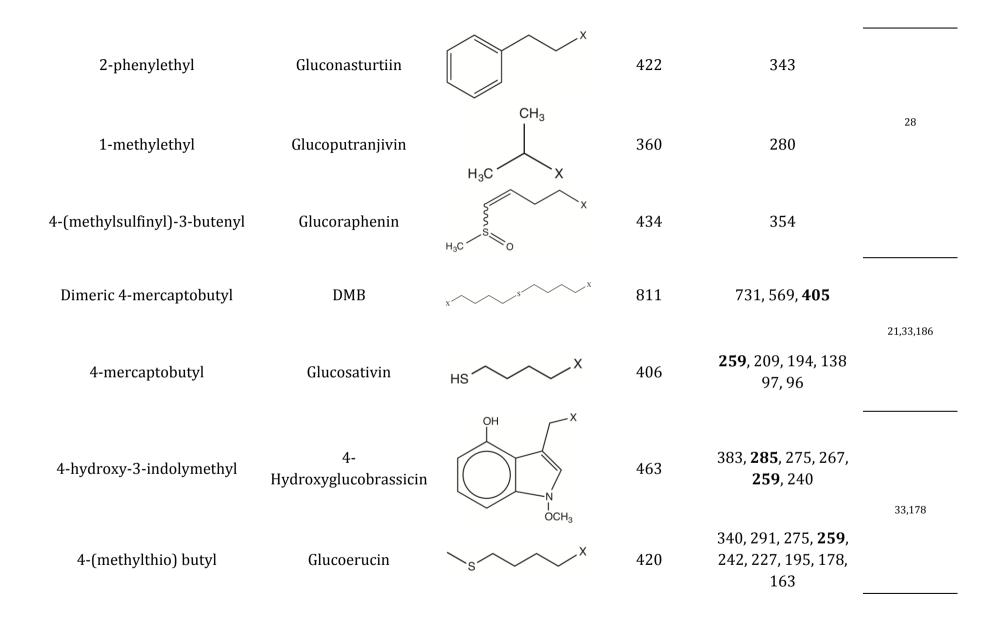
**Figure 2:** – Pathways of documented ITC action in tumorigenic cells. See Wu et al. <sup>113</sup> for a detailed review of the roles ITCs play in cancer prevention.

**Figure 3:** – The mercapturic acid pathway of ITC metabolism in the human body. After ingestion of rocket leaves glucoerucin is hydrolyzed by myrosinase to form erucin. This is released and absorbed in the ileum, where it is transported in the blood to cells around the body. ITCs initiate Phase II detoxification enzymes in this pathway, and are known to aid in cancer prevention. (Adapted from Wu et al. <sup>113</sup>).

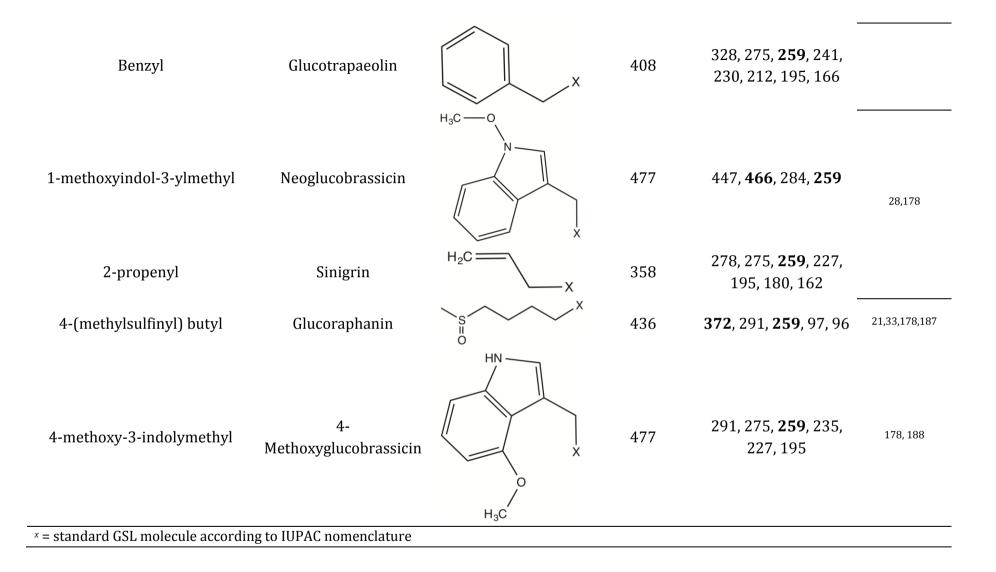
**Figure 4:** – Factors and conditions within the commercial supply chain that affect GSL and flavonol levels within rocket leaves.

R-group	Common name	R-group structure <sup>x</sup>	Mass parent ion	MS <sup>2</sup> spectrum ions (signature ions in bold)	Reference	
2-(benzoyloxy) ethyl	-		466	386	33	
3-hydroxy-5-(methylsulfinyl) pentyl	-	S S N O N N N N N N N N N N N N N N N N	482	403		
4-(β-D-glucopyranosyldisulfanyl) butyl	Diglucothiobeinin	x s x s	600	521	33,186	
5-(methylsulfinyl) pentyl	Glucoalyssin	S U O	450	371		
<i>N</i> -butyl	Dihydrogluconapin	<u> </u>	374	294		
4-phenylbutyl	Glucoamoracin	×	450	450 371		
7-(methylsulfinyl) heptyl	Glucoibarin	s s x	494	414	28	
Ethyl	Glucolepiidin	H <sub>3</sub> C X	346	266		

Table 1: - Intact Glucosinolates Identified Within Leaves Of Rocket, Eruca and Diplotaxis Species, By LC-MS (Negative Ion Mode)



4-hydroxybenzyl (R,S)-2-hydroxy-3-butenyl	Glucosinalbin Progoitrin/epiprogoitrin	HO OH X	424 388	344, 291, 275, 261, <b>259</b> , 246, 231, 228, 182 332, 308, 301, 275, 259, 210, 195, 136	33,178	
3-indolymethyl	Glucobrassicin	X	447	275, <b>259</b> , 251, 205	33,178,187	
1-methylpropyl	Glucocochlearin	×	374	294		
2-methylbutyl	Glucojiaputin	H <sub>3</sub> C CH <sub>3</sub>	388	308	85,186	
5-(methylsulfonyl) pentyl	Glucoerysihienin	H <sub>3</sub> C /// X	466	386	28,33	
3-(methylthio) propyl	Glucoiberverin	x × x	406	326, 275, <b>259</b> , 228, 145	85,178	
3-butenyl	Gluconapin	x	372	292, 275, <b>259</b> , 227, 195, 194, 176	28,178	



Flavonol compound <sup>a</sup>	Eruca <sup>p</sup>	Diplotaxis <sup>p</sup>	Mass parent ion	MS <sup>2</sup> spectrum ions (signature ion in bold)	Reference	
I 3,4'-diGlc	$\checkmark$	$\checkmark$	639	477		
I 3-Glc	$\checkmark$		477	-		
K 3-(2-Sinp-Glc)-4'-Glc	$\checkmark$		817	-		
K 3,4'-diGlc	$\checkmark$	$\checkmark$	609	-	18,33	
K 3-Glc	$\checkmark$		447	285		
Q 3-Glc	$\checkmark$		463	301		
K 3-diGlc-7-Glc	$\checkmark$		771	609		
K 3-Sinp-triGlc-7-Glc	$\checkmark$		1139	977, 771, 609, 429	33	
Q 3,4'-diGlc-3'-(6-Caf-Glc)		$\checkmark$	949	787, <b>625</b> , 463, 301		
Μ	$\checkmark$		317	151		
Q	$\checkmark$		301	151	189	
R	$\checkmark$		609	300		
(2) (2) (2, ( ( ), ( ), ( ), ( ), ( ), ( ),		$\checkmark$	1155	993, 831, 787, 669, <b>625</b> ,		
Q 3-(2-Caf-Glc)-3'-(6-Sinp-Glc)-4'-Glc				463, 301	18,25	
Q 3-(2-Mcaf-Glc)-3'-(6-Sinp-Glc)-4'-Glc		$\checkmark$	1185	1023, 817, 669, 655		
Q 3-(2-Fer-Glc)-3'-(6-Fer-Glc)-4'-Glc		$\checkmark$	1139	977, 639, 463		
Q 3-(2-Fer-Glc)-3'-(6-Sinp-Glc)-4'-Glc		$\checkmark$	1169	1007, 831, 669, 639, 463,	18,25,33	
				301		

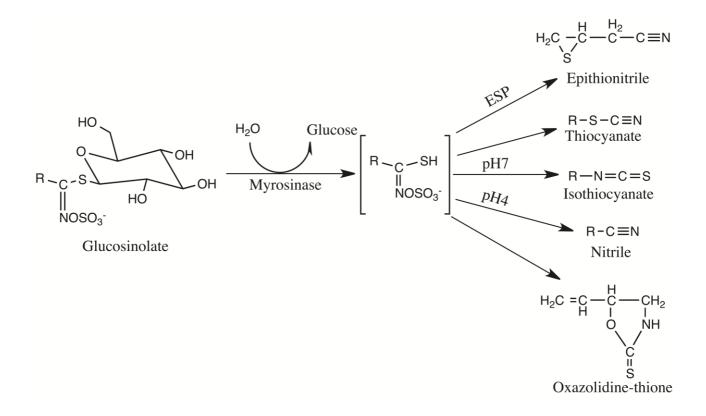
<b>Table 2:</b> – List Of Flavonol Compounds Identified In Leaves Of <i>Eruca</i> And <i>Diplotaxis</i> Species, By LC-MS (Negative Ion Mode).
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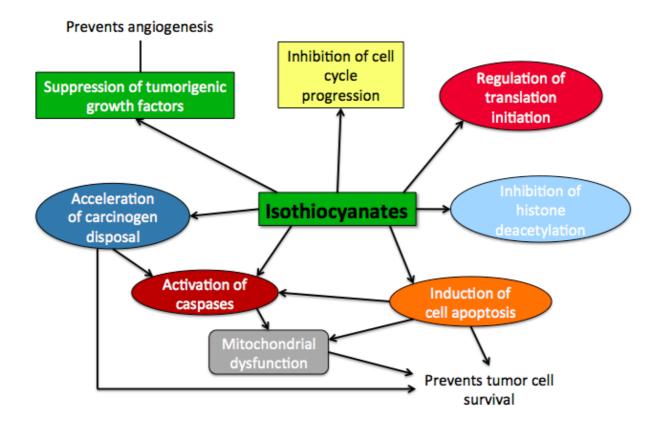
Q 3-(2-Sinp-Glc)-3'-(6-Sinp-Glc)-4'-Glc	$\checkmark$	1199		
Q 3,3',4-triGlc	$\checkmark$	787	625, 463, 301	
Q 3,4'-diGlc-3'-(6-Fer-Glc)	$\checkmark$	963	801, 639, 463, 301	18,25,33
Q 3,4'-diGlc-3'-(6-Mcaf-Glc)	$\checkmark$	979	817, 655, 463, 301	
Q 3,4'-diGlc-3'-(6- <i>p</i> .Coum-Glc)	$\checkmark$	933	771, 609, 463, 301	
Q 3,4'-diGlc-3'-(6-Sinap-Glc)	$\checkmark$	993	831, 669, 463, 301	

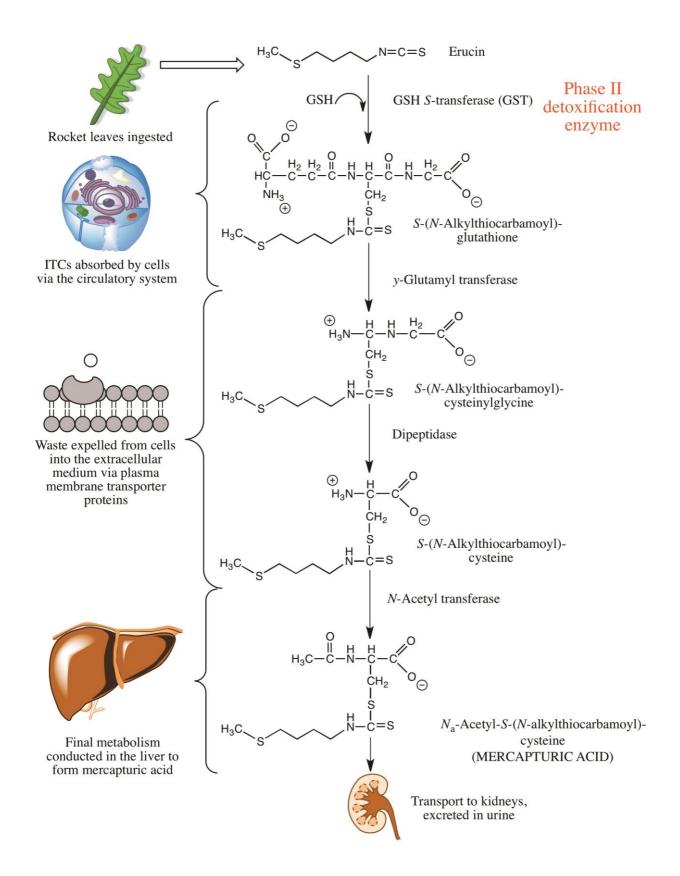
<sup>*a*</sup> = Abbreviations: Caf, caffeyol; Mcaf, methoxycaffeyol; *p*.Coum, *p*-coumaroyl; Fer, feruloyl; Sinp, sinapoyl; Glc, glucoside;

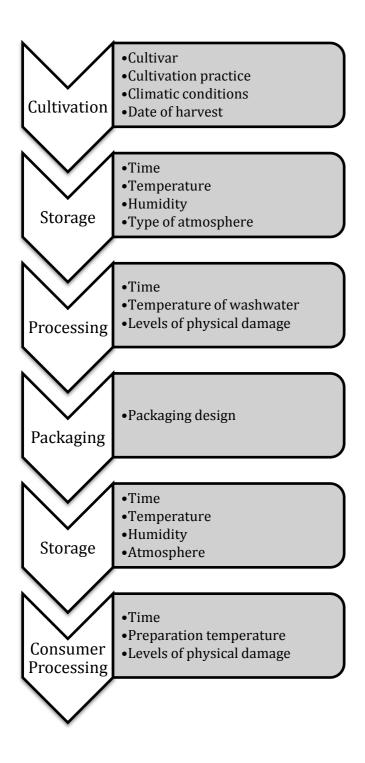
Q, quercetin; K, kaempferol; I, isorhamnetin; M, myricetin; R, rutin

 $p = \checkmark$  compound positively identified in species









## For table of contents only

