

Muddy waters: efficacious predation of container-breeding mosquitoes by a newly-described calanoid copepod across differential water clarities

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1	Muddy waters: efficacious predation of container-breeding mosquitoes by
2	a newly-described calanoid copepod across differential water clarities
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22 Abstract

Mosquito-borne diseases induce unrivalled morbidity and mortality in human 23 24 populations. In recent times, greater urbanisation has facilitated vector population expansion, 25 particularly of those which proliferate in container-style habitats. To combat increased associated disease risk, we urgently require innovative and efficacious control mechanisms to 26 27 be identified and implemented. Predatory biological control of vectorially-important mosquitoes can be effective. Despite their high prevalence in freshwater ecosystems, 28 predatory calanoid copepods have yet to be examined comprehensively for mosquito control. 29 30 Moreover, environmental context-dependencies can cause substantial variations in natural 31 enemy impacts on target populations. Accordingly, improved understanding of the effects of context-dependencies upon predatory biocontrol is needed. Here, we use functional responses 32 33 (FRs) to examine the predatory impact of a newly-described species of calanoid copepod, Lovenula raynerae, upon larval Culex pipiens prey across variations in prey supply and water 34 clarity. Using outdoor field trials, we assess the viability of L. raynerae in reducing mosquito 35 survival in container-style habitats. Lovenula raynerae displayed destabilising Type II FRs 36 towards larval mosquito prey across all water clarities tested, with overall predation rates 37 38 remaining largely unaffected across all clarity treatments. In the outdoor experiment, L. 39 raynerae applications resulted in substantial reductions in larval C. pipiens populations, with 40 close to total eradication achieved following the experimental period under higher predator 41 densities. These results demonstrate that environmental context such as water clarity may have little effect on vector control by calanoid copepods, which suggests a predatory reliance 42 43 on hydromechanical signalling. Further, for the first time, we demonstrate the applicability of 44 calanoid copepods to artificial container-style habitats where mosquitoes proliferate. 45 Therefore, our results indicate that further examination into the applicability of this species group to aid vector biocontrol strategies is warranted. 46

47 Keywords

48 biological control; predator-prey; functional response; turbidity; *Culex pipiens*; *Lovenula*49 *raynerae*

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51 1. Introduction

The effective control of mosquito-borne diseases and their vectors is of substantial 52 public health importance (Mehlhorn, 2012; Beneli and Mehlhorn, 2016; WHO, 2017). 53 54 Currently, a variety of chemical, physical, genetic and biological approaches are used to control mosquitoes (see Becker et al. 2010). However, many population management 55 approaches are associated with drawbacks which impede their sustainability (e.g. 56 Baldacchino et al. 2015). For instance, commonly-used insecticidal chemicals have caused 57 environmental pollution, and emergent effects of insecticide resistance have presented major 58 59 challenges to mosquito control strategies (e.g. Scholte et al. 2004; Ranson et al. 2016; Main et al. 2018). Mosquitoes which exploit artificial container-style habitats are of particular 60 61 public health importance due to an association with urban areas and thus high potential for 62 contact with human populations, wherein urban 'heat islands' can result in higher disease vector mosquito abundances (Townroe and Callaghan, 2014). Indeed, exploitation of human 63 environments has facilitated invasive mosquito species to radically extend their geographic 64 range (e.g. Lambrechts et al. 2010). 65

Biological control (hereafter biocontrol) provides a relatively environmentallyfriendly and economical option in vector control (Rodríguez-Pérez et al. 2012). Natural
enemies can efficaciously suppress vectorially-important mosquito populations (Marten,
1990; Marten and Reid, 2007; Baldacchino et al. 2017; Cuthbert et al. 2018a, b; but see
Thomas, 2018), and have successfully induced community-wide disease extirpations (Kay

71 and Nam, 2005; Nam et al. 2012). However, many candidate biocontrol agents remain 72 entirely unexplored, or underexploited in the context of container-style aquatic habitats where vectorially-efficient mosquitoes can proliferate en masse (e.g. Townroe and Callaghan, 73 74 2014). Biological control of larval mosquito populations by deliberate application of predatory copepod species has proven to be highly efficacious (reviewed by Marten and Reid, 75 76 2007). Presently, however, only those present within the cyclopoid order have been examined 77 and utilised for control. Yet, copepods represent a vastly extensive group of crustaceans, comprising a broad range of orders adapted to both ephemeral and perennial hydrologic 78 79 ecosystems (Dussart and Defaye, 2001). Despite previous erroneous categorisation as herbivorous, considered unable to prey upon mosquito larvae (Marten and Reid, 2007), 80 81 predatory calanoid copepods exist and can exert profound trophic impacts in aquatic 82 environments (Wasserman et al. 2016a; Dalu et al. 2016a; Cuthbert et al. 2018d). Moreover, 83 certain calanoid copepod species can be atypically large in size (e.g. Suárez-Morales et al. 2015), and can therefore handle larval mosquito stages throughout their ontogeny (Cuthbert 84 85 et al. 2018d). This contrasts to cyclopoid copepods which impart a size-refuge to larger prev (Marten and Reid 2007). Therefore, examining the efficacy of calanoid copepods towards 86 87 container-breeding mosquitoes across ranging environmental contexts is of pertinence for the applied biocontrol of mosquito-borne disease vectors. 88

Environmental context-dependencies can cause substantial variations in natural enemy impacts on target populations (e.g. Cuthbert et al. 2018a), both as a result of biotic (e.g. Alexander et al. 2013; Barrios-O'Neill et al. 2014; Wasserman et al. 2016c) and abiotic (e.g. Wasserman et al. 2016b; Cuthbert et al. 2018a, b) factors, and is thus highly relevant to biocontrol agent selection. However, the implications of these context-dependencies on the efficacy of biocontrol agents often remain poorly understood. This, in turn, reduces the capacity of practitioners to fully understand and quantify biocontrol agent impacts. As

96 vectorially-efficient mosquito species are adapted to breed in a highly variable range of 97 aquatic habitats (see Becker et al. 2010), understanding the implications of environmental context is integral to robust quantifications of biocontrol agent impacts on target mosquito 98 99 species. Further, finding biocontrol agents that are also robust to environmental variability would be desirable. In particular, water clarity is highly variable in hydrological 100 101 environments, and variations in water clarity can affect food webs though alterations of 102 predation efficacy by visual predators (e.g. van De Meutter et al. 2005; Lunt and Smee, 2015), manipulations of microhabitat structures and temperature regimes (e.g. Meysman et al. 103 104 2006; Paaijmans et al. 2017), and by directly impacting filter feeders (e.g. Rellstab and Spak, 2007), including larvae of many mosquito species. In addition, disease vector mosquitoes 105 106 have been shown be attracted to low-clarity habitats due to perceived higher nutritional loads 107 or greater depth (Ortiz-Perea and Callaghan, 2017; Cuthbert et al. 2018b), with concurrent 108 implications for mosquito abundances (e.g. Medlock and Vaux, 2014). Therefore, identifying biocontrol agents to target disease vector mosquitoes which are not impacted by turbid 109 environments is crucial for successful field applications in diverse aquatic habitats (see 110 Cuthbert et al. 2018c). 111

112 Functional responses (FRs), i.e. the per capita consumption rates of consumers with changes to resource densities (Solomon, 1949; Holling, 1959; Juliano, 2001), have been 113 114 applied extensively to quantify the resource regulation potential of consumers (e.g. Abrams, 1990; Dick et al. 2014), and can be applied to concurrently test environmental context-115 dependencies of consumer impact (e.g. South et al. 2017; Cuthbert et al. 2018a, b). Given that 116 density- and context- dependencies of *per capita* impact may affect the viability of biocontrol 117 118 agents in regulating target organisms (O'Neil, 1990; Van Driesche and Bellows, 1996; 119 Cuthbert et al. 2018a), and the regulatory efficacy of many agents is yet to be explored, here, we examine the predatory potential of Lovenula raynerae Suárez-Morales, Wasserman and 120

121 Dalu 2015, a recently described and remarkably large (4 - 5 mm) calanoid copepod species, towards larvae of the disease vector complex *Culex pipiens* in container-style environments. 122 Lovenula raynerae is a predatory ephemeral pond specialist species which hatches from 123 dormant eggs within sediment during the early stages of hydroperiod (Suárez-Morales et al. 124 2015; Wasserman et al. 2016a). Such ephemeral aquatic systems are highly varied with 125 respect to their water clarity, particularly as a result of bioturbation which can heavily impact 126 ecosystem functioning (e.g. Waterkeyn et al. 2016). Although high predatory impacts of L. 127 raynerae have recently been described upon larval mosquitoes across their ontogeny 128 129 (Cuthbert et al. 2018d), further research is required to elucidate additional contextdependencies of their impact, alongside assessments of their use in container-style habitats 130 which foster disease vector mosquitoes (Townroe and Callaghan 2014). Therefore, the 131 132 present study examines the FRs of L. raynerae towards larvae of the mosquito C. pipiens across a water clarity gradient, and also assesses the predation potential of the copepod in 133 outdoor artificial container-style habitats under varying modes of predator and prey density. 134

135 **2. Materials and methods**

136 2.1. Animal collection and rearing

Adult male and female *L. raynerae* (4 - 5 mm) were collected from an ephemeral pond in the Eastern Cape, South Africa $(33^{\circ} 10' 04.1" \text{ S } 27^{\circ} 16' 10.6" \text{ E})$ by towing a 64 µm zooplankton net through the upper water column. Copepods were transported in source water to a controlled environment (CE) room at Rhodes University, Grahamstown $(25 \text{ °C} \pm 1 \text{ °C};$ 14:10 light:dark) and housed in 25 L aquaria containing strained (200 µm) water from the collection site prior to the experiments. *Culex pipiens* complex larvae originated from egg rafts collected from artificial container-style aquatic habitats on the Rhodes University 144 campus, and were reared to the desired size class on a diet of crushed rabbit food pellets145 (Agricol, Port Eizabeth).

146 2.2. Experimental protocols

We conducted two experiments to discern the efficacy of the calanoid copepod L. 147 raynerae in mosquito control. In experiment 1, in the CE room, we quantified the effect of a 148 water clarity gradient on the predatory impact of *L. raynerae* towards larval mosquito prey. 149 Adult male L. raynerae were starved for 48 h prior to experimentation. Here, males were 150 selected for experimentation to provide standardisation of predator type, given the various 151 reproductive stages of female copepods that may influence predation rates. Functional 152 responses of copepods were constrained under three water clarity treatments, conducive with 153 the variability observed in ephemeral systems (Cuthbert, pers. obs.). Water clarity was 154 155 defined as 0%, 50% and 100% against a predefined scale using a water clarity tube (GroundTruth, Leonard) by diluting turbid water to the prescribed clarity, with each treatment 156 continuously aerated and filtered (200 μ m) prior to use. *Culex pipiens* (3.3 \pm 0.2 mm) larvae 157 were established at five prey densities (2, 4, 8, 16, 32; n = 4 per density) in 80 mL arenas of 158 5.6 cm diameter containing the appropriate clarity treatment. Once predators were added, 159 160 they were allowed to feed undisturbed for 6 h, after which they were removed and remaining prey counted to derive those killed. Controls consisted of three replicates at each density and 161 clarity treatment without predators. 162

In experiment 2, we ascertained the efficacy of *L. raynerae* in outdoor artificial container-style habitats at regulating *C. pipiens* populations. This was done in a partially shaded outdoor location within the Rhodes University campus, similar to the environments from which *C. pipiens* rafts were collected. *Culex pipiens* larvae (1.89 mm \pm 0.08) were added at two densities (50, 100), to 2 L arenas of 13.5 cm diameter, each containing 1.5 L filtered (200 μ m) aerated water from the copepod collection site and 0.3 g of crushed rabbit food pellets. Then, three predator densities were added (0, 4, 8) in a fully randomised array. We maintained copepod sex ratios of 3:1 male:female across predator treatments (i.e. 0:0, 3:1, 6:2) to minimise cannibalism (Lavens and Sorgeloos, 1996). After 72 h, the predators were removed and remaining live mosquito prey counted to derive the number eaten. We conducted at least three replicates per experimental group. Water temperatures within arenas were found to be within the 16 – 20 °C range across the duration of the experiment.

175 2.3. Statistical analyses

All statistical analyses were undertaken in R v3.4.2. (R Core Team, 2017). In 176 experiment 1, generalised linear models (GLMs) assuming a Poisson error distribution were 177 used to examine the effects of 'water clarity' and 'prey density' on raw prey consumption. 178 179 All FR analyses were undertaken within the 'frair' package in R (Pritchard et al. 2017). Logistic regression considering the proportion of prey consumed as a function of the 'prey 180 density' factor was used to infer FR types. Here, a Type II FR is determined categorically by 181 a significantly negative first order term, and a Type III FR by a significantly positive first 182 order term followed by a significantly negative second order term. We fit Rogers' random 183 predator equation to account for non-replacement of prey during the experiment (Trexler et 184 al. 1998; Juliano, 2001): 185

186
$$N_e = N_0(1 - \exp(a(N_eh - T)))$$

Eqn. 1.

- 187
- 188

where N_e is the number of prey eaten, N_0 is the initial density of prey, *a* is the attack constant, *h* is the handling time and *T* is the total experimental period. We applied the Lambert W

191 function to fit the random predator equation (Bolker, 2008). The difference (delta) method

(see Juliano, 2001) was employed to compare FR attack rates and handling times between treatments with respect to the 'water clarity' factor. We applied Bonferroni corrections to account for multiplicity of comparisons (i.e. $\alpha = 0.017$). Furthermore, we employed a nonparametric bootstrapping procedure (n = 2000) to generate 95% confidence intervals around the FR curves (see Pritchard et al. 2017).

In experiment 2, GLMs assuming a quasibinomial error distribution, as residuals were
found to be over-dispersed relative to degrees of freedom, were used to model mortality rates
with respect to the 'predator density' and 'prey density' factors. Here, we used Tukey's
comparisons *via* the 'multcomp' package in R (Hothorn et al. 2008). In all cases, nonsignificant terms and interactions were removed stepwise to obtain models with maximal
parsimony (as per Crawley, 2007).

203 **3. Results**

In experiment 1, survival in control groups was 100% and so experimental deaths of 204 larval mosquitoes were attributed to predation by copepods, which were also observed eating 205 the larvae. Overall consumption was not significantly affected by water clarity ($\gamma^2 = 1.76$, df 206 = 2, p = 0.42) but increased significantly with higher prey densities ($\chi^2 = 80.45$, df = 4, $p < 10^{-10}$ 207 0.001). The consumptive effect of 'prey density' was not dependent on the water clarity as 208 the 'water clarity × prey density' effect was not significant ($\chi^2 = 7.46$, df = 8, p = 0.49). Type 209 II FRs were detected in all water clarity treatments (Table 1; Figure 1). Functional response 210 parameters (attack rate, a; handling time, h) did not differ significantly between any water 211 clarity treatment pairs (Table 1; Figure 1; a, low – medium, z = 0.64, p = 0.52; a, medium – 212 213 high, z = 0.25, p = 0.80; a, low – high, z = 0.87, p = 0.39; h, low – medium, z = 0.55, p = 0.55, p = 0.25, 0.58; h, medium – high, z = 0.79, p = 0.43; h, low – high, z = 0.32, p = 0.75). 214

In experiment 2, overall, the presence of L. raynerae resulted in significant larval 215 mosquito reductions given that the 'predator density' factor significantly affected mortality 216 rates ($F_{2, 17} = 72.59$, p < 0.001; Figure 1). Greater mortality rates were found between all 217 incremental predator density increases (0 - 4, z = 6.69, p < 0.001; 4 - 8; z = 4.35, p < 0.001; 0218 -8, z = 9.90, p < 0.001). Significantly greater mortality rates of larval mosquito prey were 219 demonstrated under the lower prey density treatments overall ($F_{1,16} = 10.23$, p = 0.006; 220 Figure 2). There was no 'predator density × prey density' interaction ($F_{2, 14} = 1.37$, p = 0.29), 221 and so the efficacy of L. raynerae at different densities was robust to treatment variations 222 223 associated with prey density.

224 **4. Discussion**

225 Here, for the first time, we demonstrate high *per capita* predation potential of an 226 ephemeral pond specialist calanoid copepod, L. raynerae, towards vector mosquito prey irrespective of water clarity regime. Equally, in outdoor trials, we show that this species can 227 substantially reduce larval mosquito abundances in container-style habitats which frequently 228 foster vectorially-efficient mosquito species (Townroe and Callaghan, 2014). Copepods are 229 highly efficacious predatory biocontrol agents for disease vectoring mosquitoes (Marten, 230 231 1984; Marten and Reid, 2007; Cuthbert et al 2018a, b). Although biocontrol examinations have hitherto focused on cyclopoid copepods, other groups of copepods are also predatory 232 233 and thus may be of value in biocontrol strategies (Wasserman et al. 2016a; Cuthbert et al. 2018d). 234

Functional responses of the calanoid copepod *L. raynerae* were not significantly affected by variations in water clarity, either in terms of form or magnitude. In a predation context, both FR form and magnitude are powerful predictors of the interaction strengths between predators and prey (Dick et al. 2014), and may be combined with predator 239 population responses to holistically assess ecological impact (Dick et al. 2017; Cuthbert et al. 2018a, b). Three broad forms of FR have been defined (Hassell, 1978): the linear Type I, 240 hyperbolic Type II and sigmoidal Type III. Whilst Type I FRs are mechanistically restricted 241 to filter feeders (Jeschke et al. 2004), Type II FRs are conducive to high ecological impact as 242 a result of high predation pressures at low prey densities (Dick et al. 2014). Accordingly, 243 Type II FRs are particularly desirable in biocontrol contexts as they trend towards target prey 244 eradications (Cuthbert et al. 2018a). However, Type III FRs are regarded as more stabilising, 245 wherein there is a provisioning of low-density refugia for prey, driven, empirically, by 246 247 processes such as prey switching (Hassell, 1978; Cuthbert et al. 2018e).

The present study observed Type II FRs regardless of water clarity regime. Therefore, 248 L. raynerae is effectively able to locate, capture and handle prey at low densities, even in 249 250 highly turbid conditions. These results are pertinent as ephemeral aquatic ecosystems, both natural and artificial, are highly varied with respect to their water clarity regime, for instance 251 due to detritus inputs or bioturbation (Cuthbert, pers. obs.). Indeed, bioturbation associated 252 with biocontrol agents can also affect the viability of multiple management interventions in 253 aquatic habitats which target larval mosquitoes (e.g. Fry-O'Brien and Mulla, 1996). 254 255 Mosquitoes often exhibit predator avoidance behaviours when ovipositing (see Vonesh and 256 Blaustein, 2010), and water clarity can offset this avoidance behaviour under certain 257 conditions (Cuthbert et al. 2018b). For this reason, identifying predatory agents which are 258 unaffected by water clarity variations is imperative for effective biocontrol applications. Our results suggest a reliance on hydromechanical cues by this predatory calanoid copepod when 259 260 detecting and capturing mosquito prey, as opposed to visual cues, which may account for the 261 lack of overall consumptive variation between water clarity treatments. Indeed, these results corroborate with those demonstrating a lack of reliance on visual signals in cyclopoid 262

263 copepods when detecting prey across diurnal and water clarity regime shifts (e.g. Hwang and
264 Strickler, 2001; Cuthbert et al. 2018c).

Attack rates of L. raynerae were relatively unaffected by variations in water clarity, 265 and even trended towards being higher under lower water clarities. Attack rates correspond to 266 the initial slope steepness in FR curves, and thus high attack rates can be particularly 267 268 destabilising to prey populations at low prey densities. On the other hand, handling times reflect the asymptote in FR curves, and can be reciprocated to infer maximum feeding rates 269 of predators (Dick et al. 2014; Cuthbert et al. 2018a). Here, although larval mosquitoes can 270 271 exhibit high responsiveness to predatory cues (e.g. Zuharah and Lester 2011), it is feasible that low water clarity increases vulnerability to predation in larval mosquito prey, in turn 272 enhancing the capture efficiency by predators at low prey densities. Furthermore, handling 273 274 times were not significantly different across the water clarity gradient in the present study, and so maximum feeding rates were similar between treatments. Yet, handling times trended 275 towards being lowest, and thus maximum feeding rates highest, at intermediate water 276 277 clarities. Hence, we present strong and sustained destabilising predatory impacts of the calanoid copepod L. raynerae towards varying prey supplies of larval C. pipiens irrespective 278 279 of this environmental context. Importantly, although FR examinations here only considered 280 males, female L. raynerae are also voracious consumers of larval mosquitoes, with the 281 potential to kill over 5 first instar larvae per hour and the ability to handle late instar prey 282 (Cuthbert et al. 2018d). This intake rate is considerably higher than cyclopoid copepods which are often used in biological control, and which also impart a size refuge to late instar 283 284 mosquito prey (Marten and Reid, 2007).

The study also highlights that the *L. raynerae* predation efficiency observed under controlled laboratory conditions also persists in outdoor environments exposed to natural conditions, where vector mosquitoes proliferate. In outdoor experiments within container288 style habitats, L. raynerae induced substantial mortality rates in larval mosquito populations. As vectorially-important mosquitoes increasingly proliferate in such artificial, container-style 289 habitats (Townroe and Callaghan, 2014), exploring the suitability for biocontrol agents over 290 291 longer-term experiments in these environments is critical for empirical derivations of their efficacy. Indeed, these aquatic habitats can often be minute and ephemeral in nature, negating 292 the use of larger larval mosquito antagonists, such as fish (see Azevedo-Santos et al. 2016). 293 294 Our results demonstrate that, over the experimental period, higher densities of L. raynerae exhibited higher predation capacities relative to lower densities towards all prey supplies. 295 296 Thus, multiple L. raynerae conspecifics may be additive in their consumption of C. pipiens larvae. Moreover, as L. raynerae is an ephemeral pond specialist and is capable of producing 297 298 dormant, drought-resistant eggs, applications of this species to ephemeral aquatic habitats 299 which foster mosquitoes may enable predator hatching *in situ* prior to, or simultaneous with, 300 colonisation by mosquitoes. Although our results are theoretically promising in this respect, further research is required to test the efficacy of single applications of dormant eggs of L. 301 302 *raynerae* over recurrent hydroperiods, and thus over longer experimental times overall. Furthermore, examinations of prey preferences and cannibalism in L. raynerae towards 303 304 juveniles would be of value in further discerning factors that may impede their applied efficacy in biocontrol. However, it has been proposed that the most efficacious copepod 305 306 species in biocontrol are able to curtail overpopulation and growth stunting *via* cannibalism 307 of juveniles when the population becomes too high relative to the food supply (Marten and Reid, 2007). 308

In conclusion, our results suggest that calanoids and other copepod groups warrant
further consideration as biocontrol agents of disease vector mosquitoes. In particular,
ephemeral pond specialist species, such as *L. raynerae*, may be particularly promising
candidates, as they are often particularly large, develop rapidly, exhibit dormancy, and have

313 adapted to occupy relatively high trophic levels (Dalu et al. 2016a). Further, their especially large size may make L. raynerae less vulnerable to higher-order predation as compared to 314 physically smaller copepod species. Generally, biotic interactions within ephemeral aquatic 315 316 ecosystems are often poorly studied due to spatial and temporal heterogeneity (Dalu et al. 2016b), and thus these systems hold much potential for biocontrol agent exploration, as 317 demonstrated in the present study. Our results show strong and destabilising predatory 318 impacts of L. raynerae towards larvae of C. pipiens across a water clarity regime, which may 319 enable consistently high impacts upon target populations under differing environmental 320 321 conditions. Furthermore, we show efficacious predatory potential of this species in artificial container-style habitats in outdoor environments. Future research should test additional 322 environmental contexts as to their effects on biotic interaction strengths between biocontrol 323 324 agents and target organisms, and further explore the sustained potential of calanoid copepods 325 over longer hydroperiods within container-style habitats which harbour disease vector mosquitoes. 326

327 **References**

Abrams, P.A., 1990. The effects of adaptive behaviour on the type-2 functional
response. Ecology 71, 877–885.

Alexander, M.E., Dick, J.T.A., O'Connor, N.E., 2013. Trait-mediated indirect
interactions in a marine intertidal system as quantified by functional responses. Oikos 122,
1521–1531.

Azevedo-Santos, V.M., Vitule, J.R.S., Pelicice, F.M., García-Berthou, E., Simberloff,
D, 2017. Non-native fish to control *Aedes* mosquitoes: A controversial, harmful tool.
BioScience 67, 84–90.

336	Baldacchino, F., Caputo, B., Chandre, F., Drago, A., della Torre, A., Montarsi, F.,
337	Rizzoli, A. 2015. Control methods against invasive Aedes mosquitoes in Europe: a review.
338	Pest Manag. Sci. 71, 1471–1485.
339	Baldacchino, F., Bruno, M.C., Visentin, P., Blondel, K., Arnolid, D., Hauffe, H.C.,
340	Rizzoli, A., 2017. Predation efficiency of copepods against the new invasive mosquito
341	species Aedes koreicus (Diptera: Culicidae) in Italy. Eur. Zool. J. 84, 43-48.
342	Barrios-O'Neill, D., Dick, J.T.A., Emmerson, M.C., Ricciardi, A., MacIsaac, H.J.,
343	Alexander, M.E., Bovy, H.C., 2014. Fortune favours the bold: a higher predator reduces the
344	impact of a native but not an invasive intermediate predator. J. Anim. Ecol. 83, 693–701.
345	Becker, N., Petric, D., Zgomba, M., Boase, C., Madon, M.B., Dahl, C., Kaiser, A.,
346	2010. Mosquitoes and their control. Springer-Verlag, Berlin/Heidelberg.
347	Benelli, G., Mehlhorn, H. 2016. Declining malaria, rising of dengue and Zika virus:
348	insights for mosquito vector control. Parasitol. Res. 115, 1747–1754.
349	Bolker, B.M., 2008. Ecological Models and Data in R. Princeton University Press,
350	Princeton.
351	Crawley, M.J., 2007. The R book. John Wiley & Sons, Chichester.
352	Cuthbert, R.N., Dick, J.T.A., Callaghan, A., Dickey, J.W.E., 2018a. Biological control
353	agent selection under environmental change using functional responses, abundances and
354	fecundities; the Relative Control Potential (RCP) metric. Biol. Control 121, 50-57.
355	Cuthbert, R.N., Dick, J.T.A., Callaghan, A., 2018b. Interspecific variation, habitat
356	complexity and ovipositional responses modulate the efficacy of cyclopoid copepods in
357	disease vector control. Biol. Control 121, 80-87.

358	Cuthbert, R.N., Callaghan, A., Dick, J.T.A., 2018c. Dye another day: the predatory
359	impact of cyclopoid copepods on larval mosquito Culex pipiens is unaffected by dyed
360	environments. J. Vector Ecol. In press.
361	Cuthbert, R.N., Dalu, T., Wasserman, R.J., Callaghan, A., Weyl, O.L.F., Dick, J.T.A.,
362	2018d. Calanoid copepods: an overlooked tool in the control of disease vector mosquitoes. J.
363	Med. Entomol. In press.
364	Cuthbert, R.N., Dickey, J.W.E., McMorrow, C., Laverty, C., Dick, J.T.A, 2018e.
365	Resistance is futile: lack of predator switching and a preference for native prey predict the
366	success of an invasive prey species. Royal Soc. Open Sci. In press.
367	Dalu, T., Weyl, O.L.F., Froneman, P.W., Wasserman, P.W., 2016a. Trophic
368	interactions in an austral temperate ephemeral pond inferred using stable isotope analysis.
369	Hydrobiologia 768, 81–94.
370	Dalu, T., Wasserman, R.J., Dalu, M.T.B., 2016b. Agricultural intensification and
371	drought frequency increases may have landscape-level consequences for ephemeral
372	ecosystems. Glob. Change Biol. 23, 983–985.
373	Dick, J.T.A., Alexander, M.E., Jeschke, J.M., Ricciardi, A., MacIsaac, H.J.,
374	Robinson, T.B., Kumschick, S., Weyl, O.L.F., Dunn, A.M., Hatcher, M.J., Paterson, R.A.,
375	Farnsworth, K.D., Richardson, D.M., 2014. Advancing impact prediction and hypothesis
376	testing in invasion ecology using a comparative functional response approach. Biol. Invasions
377	16, 735–753.
378	Dick, J.T.A., Laverty, C., Lennon, J.J., Barrios-O'Neill, D., Mensink, P.J., Britton, R.,
379	Médoc, V., Boets, P., Alexander, M.E., Taylor, N.G., Dunn, A.M., Hatcher, M.J., Rosewarne,
380	P.J., Crookes, S., MacIsaac, H.J., Xu, M., Ricciardi, A., Wasserman, R.J., Ellender, B.R.,

Weyl, O.L.F., Lucy, F.E., Banks, P.B., Dodd, J.A., MacNeil, C., Penk, M.R., Aldridge, D.C.,

Caffrey, J.M., 2017. Invader Relative Impact Potential: a new metric to understand and
predict the ecological impacts of existing, emerging and future invasive alien species. J.
Appl. Ecol. 54, 1259–1267.

385 Dussart, B.H., Defaye, D., 2001. Introduction to the Copepoda. Guides to the
386 Identification of the Microinvertebrates of the Continental Waters of the World 16. Backhuys
387 Publishers, Leiden.

Fry-O'Brien, L.L., Mulla, M.S., 1996. Effect of tadpole shrimp, *Triops longicaudatus*,
(Notostraca: Triopsidae), on the efficacy of the microbial control agent *Bacillus thuringiensis*var. *israelensis* in experimental microcosms. J. Am. Mosq. Contr. Assoc. 12, 33–38.

Hassell, M.P., 1978. The dynamics of arthropod predator-prey systems. PrincetonUniversity Press, Princeton.

Holling, C.S., 1959. Some characteristics of simple types of predation and parasitism.
Can. Entomol. 91, 385–398.

Hothorn, T., Bretz, F., Westfall, P., 2008. Simultaneous inference in general
parametric models. Biom. J. 50, 346–363.

Hwang, J.S., Strickler, R., 2001. Can copepods differentiate prey from predator
hydromechanically? Zool. Stud. 40, 1–7

Jeschke, J.M., Kopp, M., Tollrian, R., 2004. Consumer-food systems: why type I
functional responses are exclusive to filter feeders. Biol. Rev. 79, 337–349.

401 Juliano, S.A., 2001. Non-linear curve fitting: predation and functional response

402 curves. In: Scheiner, S.M., Gurevitch, J. (Eds.), Design and Analysis of Ecological

403 Experiments. Oxford University Press, Oxford, pp. 178–196.

404	Kay, B.H., Nam, V.S., 2005. New strategy against Aedes aegypti in Vietnam. Lancet
405	365, 613–617.

406	Lambrechts, L., Scott, T.W., Gubler, D.J., 2010. Consequences of the expanding
407	global distribution of Aedes albopictus for Dengue virus transmission. PLoS Negl. Trop. Dis.
408	4, 645.
409	Lavens, P., Sorgeloos, P., 1996. Manual on the production and use of live food for
410	aquaculture (No. 361). Food and Agriculture Organization (FAO), Rome.
411	Lunt, J., Smee, D.J., 2015. Turbidity interferes with foraging success of visual but not
412	chemosensory predators. PeerJ 3, e1212.
413	Main, B.J., Everitt, A., Cornel, A.J., Hormozdiari, F., Lanzaro, G.C., 2018. Genetic
414	variation associated with increased insecticide resistance in the malaria mosquito, Anopheles
415	coluzzii. Parasit. Vectors 11, 225.
416	Marten, G.G., 1984. Impact of the copepod Mesocyclops leuckarti pilosa and the
417	green alga Kirchneriella irregularis upon larval Aedes albopictus (Diptera: Culicidae). Bull.
418	Soc. Vector Ecol. 9, 1–5.
419	Marten, G.G., 1990. Elimination of Aedes albopictus from tire piles by introducing
420	Macrocyclops albidus (Copepoda, Cyclopidae). J. Am. Mosq. Control Assoc. 6, 689-693.
421	Marten, G.G., Reid, J.W., 2007. Cyclopoid copepods. J. Am. Mosq. Control Assoc.
422	23, 65–92.
423	Medlock, J.M., Vaux, A.G.C., 2014. Colonization of a newly constructed urban
424	wetland by mosquitoes in England: implications for nuisance and vector species. J. Vector
425	Ecol. 39, 249–260.

426	Mehlhorn, H., Al-Rasheid, K.A., Al-Quraishy, S., Abdel-Ghaffar, F. 2012. Research
427	and increase of expertise in arachno-entomology are urgently needed. Parasitol. Res. 110,
428	259–265.
429	Meysman, F.J.R., Middelburg, J.J., Heip, C.H.R., 2006. Bioturbation: a fresh look at
430	Darwin's last idea. Trends Ecol. Evol. 21, 688–695.
431	Nam, V.S., Yen, N.T., Duc, H.M., Tu, T.C., Thang, V.T., Le, N.H., Le Loan, L.,
432	Huong, V.T.Q., Khanh, L.H.K., Trang, H.T.T., Lam, L.Z.Y., Kutcher, S.C., Aaskov, J.G.,
433	Jeffery, J.A.L., Ryan, P.A., Kay, B.H., 2012. Community-based control of Aedes aegypti by
434	using Mesocyclops in Southern Vietnam. Am. J. Trop. Med. Hyg. 86, 850-859.
435	O'Neil, R.J., 1990. Functional response of arthropod predators and its role in the
436	biological control of insect pests in agricultural systems. In: Dunn, P.E., Baker, R.R. (Eds.),
437	New Directions in Biological Control: Alternatives for Suppressing Agricultural Pests and
438	Diseases. Alan R Liss Inc, New York, pp. 83–96.
439	Ortiz-Perea, N., Callaghan, A., 2017. Pond dyes are Culex mosquito oviposition
440	attractants. PeerJ 5, e3361.
441	Paaijmans, K., Takken, W., Githeko, A.K., Jacobs, A.F.G., 2017. The effect of water
442	turbidity on the near-surface water temperature of larval habitats of the malaria mosquito
443	Anopheles gambiae. Int. J. Biomet. 52, 747–753.
444	Pritchard, D.W., Paterson, R.A., Bovy, H.C., Barrios-O'Neill, D., 2017. Frair: an R
445	package for fitting and comparing consumer functional responses. Method. Ecol. Evol. 8,
446	1528–1534.
447	R Core Team, 2017. R: A Language and Environment for Statistical Computing. R
448	Foundation for Statistical Computing, Vienna.

449	Ranson, H., Lissenden, N., 2016. Insecticide resistance in African Anopheles
450	mosquitoes: a worsening situation that needs urgent action to maintain malaria control.
451	Trends Parasitol. 32, 187–96.
452	Rellstab C., Spaak P., 2007. Starving with a full gut? Effect of suspended particles on
453	the fitness of Daphnia hyalina. Hydrobiol. 594, 131–139.
454	Rodríguez-Pérez, M.A., Howard, A.F.V., Reyes-Villanueva, F., 2012. Biological
455	Control of Dengue Vectors. In: Larramendy, M.L. and Soloneski, S. (Eds.) Integrated Pest
456	Management and Pest Control – Current and Future Tactics. InTech. pp. 241–270.
457	Scholte, E.J., Bart, G., Knols, J., Samson, A., Takken, W., 2004. Entomopathogenic
458	fungi for mosquito control: a review. J. Insect Sci. 4, 19.
459	Solomon, M.E., 1949. The natural control of animal populations. J. Anim. Ecol. 18,
460	1–35.
461	South, J., Dick, J.T.A., 2017. Effects of acute and chronic temperature changes on the
462	functional responses of the dogfish Scyliorhinus canicula (Linnaeus, 1758) towards
463	amphipod prey Echinogammarus marinus (Leach, 1815). Environ. Biol. Fish. 100, 1251-
464	1563.
465	Suárez-Morales, E., Wasserman, R.J., Dalu, T. 2015. A new species of Lovenula
466	Schmeil (Copepoda, Diaptomidae) from the Eastern Cape province of South Africa.
467	Crustaceana 88, 324–342.
468	Thomas, M.B. 2018. Biological control of human disease vectors: a perspective on
469	challenges and opportunities. Biocontrol 63, 61-69.

470	Townroe, S., Callaghan, A., 2014. British container breeding mosquitoes: the impact
471	of urbanisation and climate change on community composition and phenology. PLoS One 9,
472	e95325.
473	Trexler, J.C., McCulloch, C.E., Travis, J., 1988. How can the functional response best
474	be determined? Oecologia 76, 206–214.
475	Van de Meutter, F., Stoks, R., De Meester, L., 2004. Behavioural linkage of pelagic
476	prey and littoral predators: microhabitat selection by Daphnia induced by damselfly larvae.
477	Oikos 107, 265–272.
478	Van Driesche, R., Bellows, T.S., 2011. Biological Control. Springer, Berlin.
479	Vonesh, J.R., Blaustein, L., 2010. Predator-induced shifts in mosquito oviposition site
480	selection: A meta-analysis and implications for vector control. Israel J. Ecol. Evol. 56, 123-
481	139.
482	Wasserman, R.J., Alexander, M.E., Barrios-O'Neill, D., Weyl, O.L.F., Dalu, T.,
483	2016a. Using functional responses to assess predator hatching phenology implications for
484	pioneering prey in arid temporary pools. J. Plankt. Res. 38, 154–158.
485	Wasserman, R.J., Alexander, M.E., Weyl, O.J.F., Barrios-O'Neill, D., Froneman,
486	P.W., Dalu, T., 2016b. Emergent effects of structural complexity and temperature on
487	predator-prey interactions. Ecosphere 7, e01239.
488	Wasserman, R.J., Alexander, M.E., Dalu, T., Ellender, B.R., Kaiser, H., Weyl, O.L.F.,
489	2016c. Using functional responses to quantify interaction effects among predators. Funct.
490	Ecol. 30, 1988–1998.

491	Waterkeyn, A., Grillas, P., Brendonck, L., 2016. Experimental test of the ecosystem
492	impacts of the keystone predator Triops cancriformis (Branchiopoda: Notostraca) in
493	temporary ponds. Freshwater Biol. 61, 1392–1404.
494	World Health Organisation, 2017. Vector-borne diseases fact sheet.

495 http://www.who.int/mediacentre/factsheets/fs387/en/ (accessed 26 April 2018).

Zuharah, W.F., Lester, P.J., 2011. Are exotic invaders less susceptible to native
predators? A test using native and exotic mosquito species in New Zealand. Popul. Ecol. 53,
307–317.

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Tables and Figure Captions

- Table 1. First order terms generated from logistic regression of proportional prey
- 515 consumption as a function of prey density alongside parameter outputs from Rogers' random
- 516 predator equation across water clarity regimes.

Water clarity	First order term, p	<i>a</i> , <i>p</i>	<i>h</i> , <i>p</i>
0%	-0.058620 < 0.001	1.572822 < 0.001	0.104547 < 0.001
070	0.050020, < 0.001	1.572022, < 0.001	0.104547, < 0.001
50%	-0.044384, < 0.001	1.201939, < 0.001	0.087762, < 0.001
100%	-0.049398, < 0.001	1.082802, < 0.001	0.115452, < 0.001

528	Figure 1. Functional responses of Lovenula raynerae towards larval Culex pipiens prey
529	across a water clarity gradient (low, 0%; medium, 50%; high, 100%). Shaded areas represent
530	bootstrapped ($n = 2000$) confidence intervals.
531	Figure 2. Mortality rate (±SE) of larval Culex pipiens at different densities in outdoor trial
532	resulting from the presence of predatory copepod Lovenula raynerae at three densities.
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