

# *Beneficial effects of multi-species mixtures on N<sub>2</sub>O emissions from intensively managed grassland swards*

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# Beneficial effects of multi-species mixtures on N<sub>2</sub>O emissions from intensively managed grassland swards.

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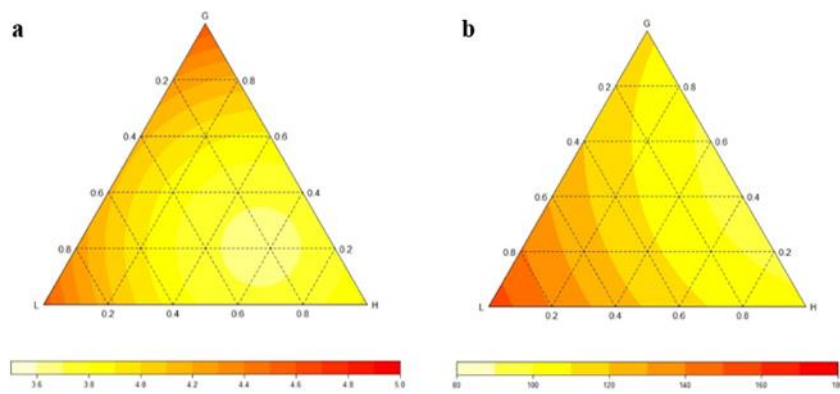
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## KEY WORDS

Grasslands, GHG emissions, nitrous oxide, yield-scaled N<sub>2</sub>O emissions, multi-species mixtures, grassland production.

## GRAPHICAL ABSTRACT



(a) N yield-scaled N<sub>2</sub>O emissions (N<sub>2</sub>O g ha<sup>-1</sup> year<sup>-1</sup> / N yield kg ha<sup>-1</sup> year<sup>-1</sup>)

(b) DM yield-scaled N<sub>2</sub>O emissions (N<sub>2</sub>O g ha<sup>-1</sup> year<sup>-1</sup> / DM yield tonne ha<sup>-1</sup> year<sup>-1</sup>)

- We assessed annual N<sub>2</sub>O emissions from field plots sown with multi-species grassland communities (1-6 species )
- N<sub>2</sub>O emissions in mixtures were best predicted from a linear combination of species' identity effects (equivalent to species' performances in monoculture), with no additional suppressive effect due to interspecific interactions.
- Based on emissions intensities, the same N yield or DM yield from the 6-species mixture and *L. perenne* monoculture could have been produced while reducing N<sub>2</sub>O losses by 41% and 24% respectively (at 150 kg ha<sup>-1</sup> year<sup>-1</sup> of nitrogen fertiliser).

[Click here to view linked References](#)

1 **Beneficial effects of multi-species mixtures on N<sub>2</sub>O emissions**  
2 **from intensively managed grassland swards.**

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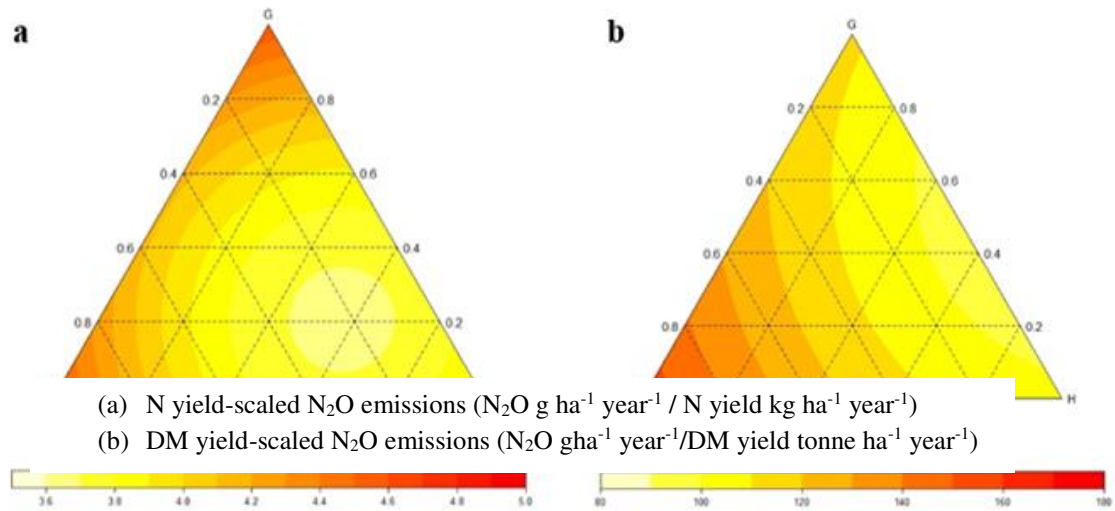
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19 **KEY WORDS**

20 Grasslands, GHG emissions, nitrous oxide, yield-scaled N<sub>2</sub>O emissions, multi-species  
21 mixtures, grassland production.

## 22 GRAPHICAL ABSTRACT



23

24

## 25 ABSTRACT

26 In a field experiment, annual N<sub>2</sub>O emissions and grassland yield were measured across  
27 different plant communities, comprising systematically varying combinations of  
28 monocultures and mixtures of three functional groups (FG): grasses (*Lolium perenne*,  
29 *Phleum pratense*), legumes (*Trifolium pratense*, *Trifolium repens*) and herbs (*Cichorium*  
30 *intybus*, *Plantago lanceolata*). Plots received 150 kg ha<sup>-1</sup> year<sup>-1</sup> N (150N), except *L.*  
31 *perenne* monocultures which received two N levels: 150N and 300N. The effect of plant  
32 diversity on N<sub>2</sub>O emissions was derived from linear combinations of species  
33 performances' in monoculture (species identity) and not from strong interactions between  
34 species in mixtures. Increasing from 150N to 300N in *L. perenne* resulted in a highly  
35 significant increase in cumulative N<sub>2</sub>O emissions from 1.39 to 3.18 kg N<sub>2</sub>O-N ha<sup>-1</sup> year<sup>-1</sup>  
36 <sup>1</sup>. Higher N<sub>2</sub>O emissions were also associated with the legume FG. Emissions intensities

37 (yield-scaled N<sub>2</sub>O emissions) from multi-species mixture communities around the equi-  
38 proportional mixture were lowered due to interactions among species. For N<sub>2</sub>O emissions  
39 scaled by nitrogen yield in forage, the 6-species mixture was significantly lower than *L.*  
40 *perenne* at both 300N and 150N. In comparison to 300N *L. perenne*, the same N yield or  
41 DM yield could have been produced with the equi-proportional 6-species mixture (150N)  
42 while reducing N<sub>2</sub>O losses by 63% and 58% respectively. Compared to 150N *L. perenne*,  
43 the same N yield or DM yield could have been produced with the 6-species mixture while  
44 reducing N<sub>2</sub>O losses by 41% and 24% respectively. Overall, this study found that multi-  
45 species grasslands can potentially reduce both N<sub>2</sub>O emissions and emissions intensities,  
46 contributing to the sustainability of grassland production.

## 47 **1. INTRODUCTION**

48 Nitrous oxide (N<sub>2</sub>O) is a potent greenhouse gas (GHG) (Ravishankara et al., 2009) with  
49 265 times the global warming potential of carbon dioxide (CO<sub>2</sub>) (IPCC, 2014). Large N<sub>2</sub>O  
50 losses result from both (N) fertiliser application to grasslands (Harty et al., 2016; Krol et  
51 al., 2020) and the N fertiliser production process itself (Wood and Cowie, 2004).  
52 Although conventional grassland systems for livestock production tend to use high levels  
53 of N fertiliser, they are not heavily reliant on imported concentrate feeds which have a  
54 high carbon footprint (O'Brien et al., 2011). Therefore, temperate grassland production  
55 systems have the potential to curtail N<sub>2</sub>O losses through the displacement of N fertiliser  
56 for symbiotically produced plant-available N.

57 Multi-species mixtures composed of grasses, legumes and herbs provide a range of  
58 agronomic and environmental benefits in grass-based production systems. These include:  
59 increased dry matter (DM) yield production (greater biomass production from species in  
60 mixtures relative to the best performing monoculture – transgressive overyielding)



61 (Nyfeler et al., 2009; Finn et al., 2013; Moloney et al., 2020), improved animal  
62 performance (for both cattle and sheep) (Cranston et al., 2015; Roca-Fernández et al.,  
63 2016; Bryant et al., 2017; Grace et al., 2019; Jerrentrup et al., 2020), increased N use  
64 efficiency (NUE) (Hooper et al., 2005, Suter et al., 2015), weed suppression (Suter et al.,  
65 2017; Connolly., 2018), and greater yield stability during drought events (Hofer et al.,  
66 2016; Haughey et al., 2019). Although these benefits are well established, less is known  
67 about how multi-species grasslands influence the soil N cycle and therefore N<sub>2</sub>O fluxes.

68 Multi-species grasslands may affect N<sub>2</sub>O fluxes in several ways (Gardiner et al., 2016;  
69 De Klein et al., 2019). Nitrous oxide is mainly lost during the soil-based processes of  
70 nitrification and denitrification (Bremner et al., 1997). Different plant species can  
71 influence these processes through differential niche occupation of the rhizosphere which  
72 can affect plant water uptake (Holtham et al., 2007) and soil gas diffusivity. These  
73 processes determine the nitrification and denitrification pathways and the final N<sub>2</sub>O:N<sub>2</sub>  
74 ratio of the denitrification process (Balaine et al., 2016). Biochemical reactions may also  
75 affect N<sub>2</sub>O production. *Plantago* species contain biological nitrification inhibition (BNI)  
76 compounds within the plant that prevent ammonium (NH<sub>4</sub><sup>+</sup>) transformation to nitrate  
77 (NO<sub>3</sub><sup>-</sup>); this results in stability of the soil mineral N pool and increased plant N uptake  
78 (Chapman et al., 2006; Cantarel et al., 2015). Legume inclusion within grassland swards  
79 and multi-species mixtures allows for reduced fertiliser application without adversely  
80 affecting yields (Egan et al., 2018). This is due to biological nitrogen fixation (BNF) and  
81 the transfer of N from legumes to non-legumes within a multi-species sward (Nyfeler et  
82 al., 2011; Pirhofer-Walzl et al., 2012). Legume inclusion in grasslands can increase N<sub>2</sub>O  
83 emissions when N fertiliser is not reduced to account for symbiotically fixed N (Hakala  
84 et al., 2012; Burchill et al., 2016; Luo et al., 2018).

85 There can be species-specific effects of plants on N<sub>2</sub>O emissions. A laboratory incubation  
86 by Abalos et al. (2014) quantified the N<sub>2</sub>O fluxes from mixtures with up to four grass  
87 species including *L. perenne*, *Festuca arundinacea*, *P. pratense* and *Poa trivialis*. No  
88 relationship was found between plant species richness and N<sub>2</sub>O emissions; however, there  
89 was a significant reduction in N<sub>2</sub>O emissions when certain plant species were combined.  
90 In a field study, Luo et al. (2018) applied cattle urine and compared the N<sub>2</sub>O fluxes from  
91 monocultures of *P. lanceolata* and *Medicago sativa* with a *T. repens* and *L. perenne*  
92 mixture. Despite seasonal variation, *P. lanceolata* had lower N<sub>2</sub>O emissions than *L.*  
93 *perenne* throughout the year. Although these studies showed potential for multi-species  
94 swards to reduce N<sub>2</sub>O emissions, the various experiments were either not monitored on  
95 an annual basis, across a range of plant communities or did not apply different N fertiliser  
96 levels. These three considerations are each important to properly understand the  
97 application of multi-species swards in livestock production systems.

98 Fuchs et al. (2020) modelled the N<sub>2</sub>O mitigation potential and productivity of various  
99 combinations of legume proportions and fertilizer rates for five temperate grassland sites  
100 using two different biogeochemical models. They recommended further study of the  
101 effect of clover proportions ranging from 30–50% receiving  $\leq 150$  kg N ha<sup>-1</sup> yr<sup>-1</sup> input,  
102 as these were identified as best-bet climate smart agricultural practices. Our study directly  
103 responds to those research recommendations. In a year-long experiment, we investigated  
104 N<sub>2</sub>O fluxes from six different forage types including two species from each of three FGs:  
105 grasses (*L. perenne*, *P. pratense*), legumes (*T. pratense*, *T. repens*) and herbs (*C. intybus*  
106 and *P. lanceolata*) in monocultures and mixtures of systematically varying proportions.  
107 In addition, emission intensity was calculated for all treatments (van Groenigen et al.,  
108 2010) as N yield-scaled N<sub>2</sub>O emissions (N<sub>2</sub>O-N g ha<sup>-1</sup> year<sup>-1</sup>/ kg N yield ha<sup>-1</sup> year<sup>-1</sup>) and  
109 DM yield-scaled N<sub>2</sub>O emissions (N<sub>2</sub>O-N g ha<sup>-1</sup> year<sup>-1</sup>/ tonne DM yield ha<sup>-1</sup> year<sup>-1</sup>). The

110 experimental design and analyses allowed quantification of species identity effects and  
111 species interaction effects on each of the three responses (N<sub>2</sub>O emissions, N yield-scaled  
112 emissions and DM yield-scaled emissions). There was also a 300 kg N ha<sup>-1</sup> year<sup>-1</sup> *L.*  
113 *perenne* treatment (300N *L. perenne*) to allow for a comparison between all treatments  
114 and conventional agricultural practice. The specific aims of this study were to:

115 1) Investigate the effect of systematically varying species and FG proportions within  
116 grassland communities on annual N<sub>2</sub>O emissions, N yield-scaled N<sub>2</sub>O emissions and DM  
117 yield-scaled N<sub>2</sub>O emissions.

118 2) Compare annual N<sub>2</sub>O emissions and yield-scaled N<sub>2</sub>O emissions from 150N forage  
119 communities with the 300N *L. perenne* community.

## 120 **2. MATERIALS & METHODS**

### 121 ***2.1 Experimental site***

122 The year-long field experiment took place at Teagasc, Johnstown Castle, Co. Wexford,  
123 Ireland 52°18'27 N between March 2018 – March 2019. The climate is temperate  
124 maritime and meteorological data (precipitation, air and soil temperature) was recorded  
125 at the Johnstown Castle weather station. The soil type at the field site was a stagnic brown  
126 podzolic. Soil texture was sandy loam, pH was 5.7 and the average bulk density of the  
127 plots on the trial site at 5-10 cm depth was 1.35 g cm<sup>-3</sup>.

### 128 ***2.2 Experimental design***

129 The experimental site was treated with herbicide, ploughed, and reseeded in spring 2017.  
130 The experiment followed a simplex design (Scheffe, 1963) for use in conjunction with  
131 the statistical modelling described in Section 2.5. Experimental plots, each measuring 5  
132 m x 7 m were sown with grassland communities (*Appendix A and D*) comprising one to

133 six species that systematically varied FG composition and relative abundance. The six  
134 species comprised two species from each of three FGs: two grasses (*L. perenne* and *P.*  
135 *pratense*), two clovers (*T. repens* and *T. pratense*) and two deep-rooting herbs (*C. intybus*  
136 and *P. lanceolata*). There were 20 different communities with between one to four  
137 replicates per treatment (*Appendix B*) resulting in 43 experimental plots in total. Each  
138 main plot was divided into two 5 m x 3.5 m sub plots, and two water supply treatments  
139 were applied at random to the two halves. One split plot (randomly chosen) received  
140 natural water supply over the year ('rain fed'), while a two-month summer drought was  
141 simulated on the other half, using rainout shelters ('drought'). Here, we only report the  
142 measured N<sub>2</sub>O emissions from the rain fed sub plots. Due to the natural drought  
143 conditions during the summer of 2018, the rain fed sub plots were irrigated on three  
144 occasions with 30mm of water, to match historical rainfall records (Met Éireann, 2020).  
145 Fertiliser N application was divided into five applications of varying rate from March-  
146 September 2018 (*Appendix C*). Maintenance levels of P and K fertilisers were applied in  
147 line with soil test recommendations at the beginning of the growing season. Calcium  
148 ammonium nitrate (CAN) fertiliser was applied at rates of 150 kg ha<sup>-1</sup> year<sup>-1</sup> (150N;  
149 communities 1-19, *Appendix B*) and 300 kg ha<sup>-1</sup> year<sup>-1</sup> (300N *L. perenne*; community 20,  
150 *Appendix B*).

### 151 ***2.3 Nitrous oxide measurements***

152 Nitrous oxide emissions were monitored from 13<sup>th</sup> March 2018 – 21<sup>st</sup> March 2019. To  
153 capture fertiliser-induced effects on N<sub>2</sub>O fluxes, a high resolution N<sub>2</sub>O sampling strategy  
154 was put in place for six months (March to September 2018) in order to coincide with  
155 fertiliser application - the time that emissions were expected to be highest. Sampling took  
156 place four days a week for two weeks immediately following each fertiliser application,  
157 two days a week in the next two weeks (weeks three and four) and once per week up until

158 the next fertiliser application date. High-resolution N<sub>2</sub>O sampling was followed by six  
159 months of low-resolution sampling at a frequency of once a month (October 2018 to  
160 March 2019). The less intensive sampling approach is reflective of the low N<sub>2</sub>O fluxes  
161 expected during this period due to a combination of no N fertiliser application and low  
162 soil temperature (Maire et al., 2020).

163 Nitrous oxide was measured using static chamber methodology (De Klein and Harvey,  
164 2012), with a single chamber placed in each plot giving a total of 43 chambers. Chambers  
165 consisted of square, stainless steel collars 40 cm (length) × 40 cm (breadth) × 10cm  
166 (height) lined with a neoprene strip and inserted to 5 cm soil depth with matching steel  
167 covers creating an approximately 16 litre headspace. A 10 kg weight was placed on top  
168 of the covers at sampling times to ensure an airtight headspace for an enclosure time of  
169 40 minutes. A 10 ml air sample was removed through a 16 mm rubber septum using a 10  
170 ml polypropylene syringe and hypodermic needle. The syringe was filled and emptied  
171 twice within the chamber to mix the headspace air prior to sampling. The gas samples  
172 were injected into pre-evacuated (-1,000 mbar) 7 ml screw-cap septum glass vials. Gas  
173 samples were taken from each chamber at 0, 20 and 40 minutes to measure N<sub>2</sub>O  
174 concentration over time. Sampling events took place between the hours of 10:00 and  
175 13:00 to obtain measurements representative of the average hourly flux of the day (De  
176 Klein & Harvey, 2012). Nitrous oxide concentrations were analysed using a gas  
177 chromatograph (GC, Varian CP 3800 GC, Varian, USA) fitted with an electron capture  
178 detector using high-purity helium as the carrier gas. Quality control N<sub>2</sub>O standards, which  
179 were representative of the upper N<sub>2</sub>O concentration limit expected, were analysed  
180 alongside N<sub>2</sub>O field samples. Linear regression of the increase in N<sub>2</sub>O gas concentrations  
181 over time (0, 20 and 40 minutes) was used to calculate daily fluxes (g N ha<sup>-1</sup> day<sup>-1</sup>). A  
182 single annual cumulative N<sub>2</sub>O value was calculated per plot by integration of daily fluxes

183 and linear interpolation between measurements (Burchill et al., 2014; De Klein and  
184 Harvey, 2012). Yield scaled-N<sub>2</sub>O emissions (van Groenigen et al., 2010; Sanz-Cobena et  
185 al., 2014) were calculated by dividing annual cumulative g N<sub>2</sub>O-N (g ha<sup>-1</sup> year<sup>-1</sup>) by 1)  
186 aboveground N yield (kg ha<sup>-1</sup> year<sup>-1</sup>) and 2) DM yield (tonnes ha<sup>-1</sup> year<sup>-1</sup>).

#### 187 *2.4 Ancillary measurements*

188 An area of the experimental plots was designated for ancillary measurements. A  
189 meteorological station was located approximately 500 m from the experimental site. Air  
190 temperature and atmospheric pressure were noted at each N<sub>2</sub>O sampling occasion along  
191 with volumetric soil water content using a Theta probe (type ML2; Delta-T Devices,  
192 Cambridge, UK). Soil moisture measurements were used to calculate the water filled pore  
193 space % (WFPS) (Equation 1):

194 **Equation 1.** Where SWC = volumetric soil water content, BD = bulk density and PD =  
195 particle density (Fichtner et al., 2019).

$$196 \quad WFPS \% = \frac{SWC}{1 - \frac{BD}{PD}} \times 100$$

#### 197 *2.5 Data analyses*

198 The three response variables (y) were: N<sub>2</sub>O-N emissions (kg ha<sup>-1</sup> year<sup>-1</sup>), N yield-scaled  
199 N<sub>2</sub>O emissions (N<sub>2</sub>O-N g ha<sup>-1</sup> year<sup>-1</sup>/ kg N yield ha<sup>-1</sup> year<sup>-1</sup>) and DM yield-scaled N<sub>2</sub>O  
200 emissions (N<sub>2</sub>O-N g ha<sup>-1</sup> year<sup>-1</sup>/ tonne DM yield ha<sup>-1</sup> year<sup>-1</sup>). Using the regression-based  
201 Diversity-Interactions modelling approach (Kirwan et al., 2009), we regressed responses  
202 on the sown proportional contributions of the six species as follows:

203 **Equation 2.** Regression model equation

204

$$y = \sum_{i=1}^6 \beta_i P_i + \beta_7 P_{Lp300N} + \delta \sum_{\substack{i,j=1 \\ i < j}}^6 P_i P_j + \varepsilon$$

205

Where  $y$  is the response variable (model fitted separately to each of our three responses),

206

$P_i$  represents the sown proportion of a species in a community (for  $i$ : 1 = *L. perenne*, 2 =

207

*P. pratense*, 3 = *T. pratense*, 4 = *T. repens*, 5 = *C. intybus* and 6 = *P. lanceolata*). The  $\beta_1$

208

to  $\beta_6$  coefficient are the identity effects of each species (under 150N fertiliser); if  $P_i = 1$ ,

209

the  $\beta_i$  coefficient is the expected monoculture response of species  $i$ , while if  $P_i < 1$ , the

210

expected contribution of that species to the mixture is  $\beta_i P_i$ . An extra term ( $\beta_7$ ) was

211

included for the 300N *L. perenne* monoculture plots ( $P_{Lp300N} = 1$  for these plots and 0

212

otherwise). Equation 2 assumes that all pairs of species interact in the same way (captured

213

by the coefficient  $\delta$ ). We tested various forms of the interactions, including no interaction

214

effects and whether pairwise interactions were determined by FG membership (Kirwan

215

et al., 2009). The error term  $\varepsilon$  was initially assumed to be normally distributed with zero

216

mean and constant variance  $\sigma^2$ . However, exploratory analysis indicated that responses

217

from plots with 100% legume were considerably more variable than all other plots,

218

therefore we assumed that the error was normally distributed with zero mean and with

219

two variance terms depending on the sown proportion of legume (100% or <100%). The

220

Diversity-Interactions modelling approach allows prediction of the response for a wide

221

range of communities from this six-species pool, based on the relative proportions of the

222

component species. The overall response is based on the linear combinations of the

223

identity effects, plus the sum of the interaction effects as required. Thus, for example, for

224

a 50:50 grass-legume mixture of *L. perenne* and *T. pratense*, the expected response is

225

$(\beta_1)0.5 + (\beta_3)0.5 + (\delta)(0.5*0.5)$ . We predicted from the final fitted model to assess the

226

effects on our three response variables across the monocultures and selected communities,

227 which included the 6-species mixture and 300N *L. perenne* monoculture. The analysis  
228 was performed using SAS software version the software package SAS version 9.4 (SAS  
229 Institute, Cary, North Carolina, USA).

### 230 **3. RESULTS**

#### 231 *3.1 Climatic conditions*

232 The highest average daily temperature (Fig. 1) recorded at the field site was in July at  
233 20.4°C. The lowest daily average temperature was 18<sup>th</sup> March 2018 at 0.1°C. These are  
234 in contrast to long-term climatic averages recorded at the Rosslare Co. Wexford station  
235 (10 km away). Between 1978 and 2007, on average the highest daily temperature was  
236 13.1°C and the lowest was 8.1°C. The long term mean annual rainfall for Johnstown  
237 Castle was 905.5 mm, with 49.9 mm for July, whereas the total annual rainfall for 2018-  
238 2019 (Fig. 1) was 1089.4 mm, with the average monthly rainfall for July 2018 being 1.7  
239 mm (not including irrigation). The WFPS of soil at the experimental site averaged 48%  
240 over the experimental year (Fig. 2). Following high levels of precipitation in early 2018  
241 (Fig. 1), the WFPS stayed at ~70% until early May, whereas during June, July and August  
242 the WFPS declined to 20-30%.

243 **Fig. 1. Precipitation (mm) and temperature (°C)**

#### 244 *3.2 Nitrous oxide emissions*

##### 245 *3.2.1 Seasonal patterns in N<sub>2</sub>O emissions*

246 High N<sub>2</sub>O fluxes were measured in April 2018, coinciding with high soil WFPS during  
247 March 2018 (Fig. 2). Conversely, low daily average N<sub>2</sub>O fluxes were recorded during the  
248 drought period from June-August 2018 (Fig. 2). The highest daily average N<sub>2</sub>O flux  
249 recorded (cumulative flux/number of days within the period) during the experimental year



250 was from 300N *L. perenne* at 17.85 g ha<sup>-1</sup> day<sup>-1</sup> and the lowest was from *P. lanceolata* at  
251 4.07 g N<sub>2</sub>O ha<sup>-1</sup> day<sup>-1</sup>. The 150N *L. perenne* had a daily average N<sub>2</sub>O flux of 5.40 g ha<sup>-1</sup>  
252 day<sup>-1</sup>. The highest individual N<sub>2</sub>O measurement was from 300N *L. perenne* at 112 g ha<sup>-1</sup>  
253 day<sup>-1</sup> (Fig. 2) on 18<sup>th</sup> April. From April to May, (Fig. 2) the daily average N<sub>2</sub>O flux of  
254 300N *L. perenne* was nine times higher than the 6-species mixture and five times greater  
255 than 150N *L. perenne*. No N fertiliser was applied after September 2018 (in line with the  
256 Nitrates Directive), resulting in little to no N<sub>2</sub>O fluxes during the autumn and winter  
257 period, except for the legume monocultures. Both clover species continued to produce  
258 N<sub>2</sub>O emissions in the autumn/winter period (Fig. 2) with 36% of legume N<sub>2</sub>O emissions  
259 occurring from August 2018 to January 2019, a time usually associated with low  
260 emissions.

261 **Fig. 2.** Nitrous oxide emissions and corresponding water filled pore space (WFPS %)

### 262 **3.2.2 Effects of plant diversity on cumulative N<sub>2</sub>O emissions**

263 According to model comparisons, the best model for the N<sub>2</sub>O emissions included species  
264 identity effects for each species and a term for 300N *L. perenne*, but no effects of species  
265 interactions were detected (model coefficient estimates shown in Table 1, first column).  
266 Thus, the effects of plant diversity on N<sub>2</sub>O emissions were derived from species identity  
267 effects and their linear combination in mixtures, rather than from synergistic or  
268 antagonistic species interaction effects in mixtures (as shown in equation 2, but with the  
269 last term involving  $\delta$  omitted). There was no significant difference between the 6-species  
270 mixture and any of the 150N monocultures, with the exception of the *C. intybus*  
271 monoculture (Fig. 3). The N<sub>2</sub>O emissions from 300N *L. perenne* (3.18 kg ha<sup>-1</sup> year<sup>-1</sup>) were  
272 over twice that of the 6-species mixture (1.52 kg ha<sup>-1</sup> year<sup>-1</sup>) (Table 1, Fig. 3).

273 **Table 1.** (a) Coefficient estimates  $\pm$  standard errors for the identity effects ( $\beta$ ) and  
274 interaction estimates ( $\delta$ )

275 **Fig. 3.** Comparison of predicted N<sub>2</sub>O-N emissions

276 The highest N<sub>2</sub>O emissions were from 300N *L. perenne* (significantly higher than all other  
277 treatments). Increasing fertiliser application to a *L. perenne* monoculture from 150N to  
278 300N increased (P < 0.001) cumulative N<sub>2</sub>O emissions from 1.39 to 3.18 N<sub>2</sub>O-N kg ha<sup>-1</sup>  
279 year<sup>-1</sup> (Table 1, Fig. 3). N<sub>2</sub>O emissions from 300N *L. perenne* were nearly three times  
280 higher than those from the *C. intybus* monoculture, the latter having the lowest estimated  
281 annual emissions at 1.1 kg ha<sup>-1</sup> year<sup>-1</sup>.

282 The ternary diagram (Fig. 4a) displays how variation in FG proportion (grass, herb and  
283 legume) affected the predicted annual N<sub>2</sub>O emissions; higher N<sub>2</sub>O emissions resulted  
284 from increased legume proportion while lower emissions were associated with  
285 communities dominated by grasses and/or herbs. The annual N<sub>2</sub>O emissions from the  
286 community comprising 100% legume FG (50% *T. pratense* and 50% *T. repens*) were  
287 significantly higher (P = 0.033 and P = 0.007) than those from the equi-proportional  
288 community of either the grass (150N *L. perenne* and *P. pratense*) or herb (*C. intybus* and  
289 *P. lanceolata*) FGs, respectively (Fig. 4b).

290 **Fig. 4 a)** Predicted annual N<sub>2</sub>O emissions (N<sub>2</sub>O-N kg ha<sup>-1</sup> year<sup>-1</sup>) in response to variation  
291 within grassland communities. **b)** Predicted annual N<sub>2</sub>O emissions (N<sub>2</sub>O-N kg ha<sup>-1</sup> year<sup>-1</sup>  
292 from each FG: grass, legume and herb.

### 293 **3.3 Yield-scaled N<sub>2</sub>O analyses (emissions intensity)**

294 For 2018, the average N-yield in harvested forage of the 6-species mixture, 150N *L.*  
295 *perenne* and 300N *L. perenne* was 40.5 kg ha<sup>-1</sup> year<sup>-1</sup>, 19.9 kg ha<sup>-1</sup> year<sup>-1</sup> and 28.5 kg ha<sup>-1</sup>

296  $\text{year}^{-1}$  respectively. The 6-species mixture produced an average DM yield of 12.4 tonnes  
297  $\text{DM ha}^{-1} \text{ year}^{-1}$ , and *L. perenne* at 150N and 300N produced 9.2 and 10.7 tonnes  $\text{DM ha}^{-1}$   
298  $\text{year}^{-1}$  respectively (Grange et al., in review). This yield data was combined with  $\text{N}_2\text{O}$   
299 data to calculate two measures of yield-scaled  $\text{N}_2\text{O}$  emissions; as outlined in section 2,  
300 the two measures of emission intensity analysed were N yield-scaled  $\text{N}_2\text{O}$  emissions  
301 ( $\text{N}_2\text{O-N g ha}^{-1} \text{ year}^{-1} / \text{kg N yield ha}^{-1} \text{ year}^{-1}$ ) and DM yield-scaled  $\text{N}_2\text{O}$  emissions ( $\text{N}_2\text{O-}$   
302  $\text{N g ha}^{-1} \text{ year}^{-1} / \text{tonne DM yield ha}^{-1} \text{ year}^{-1}$ ).

303 For both measures of emission intensity, the best model included species identity effects  
304 for each species, and a negative average pairwise interaction effect that resulted in an  
305 additional suppressive effect on yield-scaled  $\text{N}_2\text{O}$  emissions in mixtures (as in equation  
306 2, with model estimates in Table 1). The suppressive interaction term was only borderline  
307 significant for the DM yield-scaled emissions ( $P = 0.056$ ), however, diagnostic analysis  
308 of the model with and without the interaction term indicated a superior fit when the  
309 interaction term was included and it was kept in the final model. Emissions from the 6-  
310 species mixture were lower than the mean of the six 150N monocultures for N yield-  
311 scaled emissions ( $P = 0.012$ ) and there was a similar indication for DM yield-scaled  
312 emissions ( $P = 0.056$ ), demonstrating that increasing species diversity in multi-species  
313 mixtures suppressed yield-scaled  $\text{N}_2\text{O}$  emissions (Fig. 5a and 5b). The 300N *L. perenne*  
314 treatment had higher N yield-scaled emissions (Fig. 5a) than all other 150N communities.  
315 This result was similar for DM yield-scaled emissions (Fig. 5b), with the exception of  
316 both legume monocultures.

317 As the proportions of grasses, legumes and herbs change, communities with high  
318 proportions of legumes and/or grasses showed an increase in N yield-scaled  $\text{N}_2\text{O}$   
319 emissions (Fig. 6a). Communities with high legume proportion showed an increase in  
320 DM yield-scaled  $\text{N}_2\text{O}$  emissions (Fig. 6b) compared with herbs and grasses.

321 **Fig. 5. a and b** Comparison of yield-scaled N<sub>2</sub>O emissions.

322 **Fig. 6. a and b** Estimated emission intensity analyses (ternary diagrams).

## 323 **4. DISCUSSION**

### 324 *4.1 N<sub>2</sub>O emissions*

325 Nitrous oxide emissions from multi-species mixtures were best explained as a linear  
326 combination of the identity effects, showing that there was no net synergy or antagonism  
327 attributable to interspecific interactions. Given the large magnitude of differences among  
328 the species' identity effects, emissions from mixture compositions at and around the equi-  
329 proportional 6-species mixture tended to be considerably lower than those from the  
330 highest-emitting communities (Fig. 4), e.g. those dominated by legumes. There were no  
331 net synergistic or antagonistic effects due to interspecific interactions; nevertheless, there  
332 was still a benefit from multi-species mixtures through a reduction in the proportions of  
333 higher-emitting species in the mixture, i.e., legumes (Fig. 4a and b).

334 Our study highlights the potential for the multi-functional benefits associated with diverse  
335 grassland communities (e.g., see Introduction) to be gained without an associated increase  
336 in N<sub>2</sub>O emissions. Comparing N<sub>2</sub>O emissions from the 6-species mixture and 300N *L.*  
337 *perenne*, greater DM yield and total N yield can be obtained from the six-species mixture  
338 while halving N<sub>2</sub>O emissions (1.52 vs 3.18 N<sub>2</sub>O-N kg ha<sup>-1</sup> year<sup>-1</sup>) (Table 1). The strong  
339 effect of higher fertilizer application (300N vs 150N) on N<sub>2</sub>O emissions probably reflects  
340 non-plant uptake of excess mineral N in the soil, and these results align with the previous  
341 findings of Harty et al., (2016), Krol et al. (2016) and Cardenas et al (2020). Our results  
342 reiterate the reduced nitrogen use efficiency associated with conventional grassland *L.*  
343 *perenne* monocultures receiving high N fertiliser application.

344 Overall, increasing the proportion of both grasses and herbs within a grassland  
345 community resulted in lower N<sub>2</sub>O emissions than in legume-dominated communities  
346 (Fig. 4a). When comparing FGs, legumes had significantly higher N<sub>2</sub>O emissions than  
347 both herb and grass communities (Fig. 4b). When looking at selected monocultures, there  
348 was no significant difference in N<sub>2</sub>O emissions between *P. lanceolata* and 150N *L.*  
349 *perenne* (Fig. 3). This is surprising given that many studies have found that swards  
350 dominated by *Plantago* species can directly reduce N<sub>2</sub>O emissions in comparison with *L.*  
351 *perenne* monocultures (Gardiner et al., 2018; Luo et al., 2018). Bracken et al. (2020)  
352 found evidence that *P. lanceolata* potentially inhibits nitrification when included in a  
353 mixed sward. In contrast to aforementioned studies, the lowest N<sub>2</sub>O emissions in our  
354 experiment were from the *C. intybus* monoculture rather than *P. lanceolata*.

355 When looking at annual N<sub>2</sub>O emissions from selected individual monocultures (Fig. 3),  
356 each of *T. repens* and *T. pratense* did not have significantly higher N<sub>2</sub>O emissions than  
357 other monocultures (with the exception of *T. pratense* being higher than *C. intybus* (Fig.  
358 3)). This is strongly related to the much higher variability associated with legume  
359 monocultures in this experiment, compared to the other species (Table 1). If this is a  
360 general occurrence, legume monocultures may need more replication to have sufficient  
361 power to test their difference from other monocultures. When averaging over the species-  
362 to-species variation within each functional group, an effect of the legume FG on N<sub>2</sub>O  
363 emissions of N<sub>2</sub>O was evident (Fig. 4a, b). Many studies report high N<sub>2</sub>O emissions from  
364 legume-based pastures due to the accumulation of nitrate following mineralization of  
365 biologically fixed organic N (e.g., Dalal et al., 2003; Burchill et al., 2014).

366 Greater variation in N<sub>2</sub>O emissions from legumes may stem from the high levels of  
367 variation associated with BNF (and therefore N<sub>2</sub>O emissions from legume stands)  
368 (Rochette et al., 2004; Unkovich et al., 2008; Evers., 2011; Nyfeler et al., 2011;). Across

369 studies, the rate of BNF can range from 100 to 380 kg N ha<sup>-1</sup> year<sup>-1</sup> in northern temperate  
370 pastures and is dependent on multiple factors including environmental conditions,  
371 grassland management practices and legume species/cultivar type and proportion  
372 (Ledgard and Steele 1992; Hansen and Vinther 2001; Fox et al., 2019). Nitrous oxide  
373 fluxes associated with legumes can be attributed to N release from root exudates and from  
374 crop residue decomposition, rather than from the BNF process itself (Rochette and  
375 Jansen., 2005). The latter results were pivotal in the removal of the BNF process from the  
376 Intergovernmental Panel on Climate Change (IPCC) N<sub>2</sub>O inventory methodology.  
377 Legume inclusion within a grassland community allows for N addition into the soil  
378 system by BNF, lowering reliance on N fertilizer application without compromising on  
379 yield (Egan et al., 2018). Legume residues can improve the quality and quantity of soil  
380 organic matter over time, providing benefits for following crops (termed 'legacy effect').  
381 Fox et al. (2019) assessed legacy effects over a range of legume proportions and N  
382 application levels, and found maximum legacy benefits on a *L. multiflorum* crop from a  
383 prior grassland ley comprising 50% legume proportion and receiving 150N. Our study  
384 should be considered when using legumes as ley cover crops as environmental benefits  
385 of N addition to soil (i.e., less fertilizer N requirement) may be compromised by high N<sub>2</sub>O  
386 losses.

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#### 388 ***4.2 Yield-scaled N<sub>2</sub>O emissions (emissions intensity)***

389 The effects of multi-species swards were more pronounced when considering yield-scaled  
390 N<sub>2</sub>O emissions (expressed as either N<sub>2</sub>O-N g ha<sup>-1</sup> year<sup>-1</sup> / N yield kg ha<sup>-1</sup> year<sup>-1</sup> or N<sub>2</sub>O-N  
391 g ha<sup>-1</sup> year<sup>-1</sup> / DM yield tonne ha<sup>-1</sup> year<sup>-1</sup>). Looking at both responses, the performance of  
392 mixtures was best explained as a linear combination of the identity effects, and an

393 additional antagonistic interaction between species that acted to suppress emissions  
394 intensity. Thus, compared to the yield-scaled N<sub>2</sub>O emissions predicted from the average  
395 of the six monocultures, the yield-scaled N<sub>2</sub>O emissions of the 6-species mixture were  
396 29.1 % lower (P = 0.012) and 24.9% lower (P = 0.056) for the N- and DM yield-scaled  
397 measures respectively (Table 1). Given the differences among the species, mixture  
398 compositions at and around the 6-species mixture tended to have considerably lower  
399 yield-scaled N<sub>2</sub>O emissions (Fig. 6a and b). As both yield-scaled N<sub>2</sub>O responses had the  
400 same numerator (N<sub>2</sub>O emissions) the significant diversity effect must be related to a  
401 strong effect of plant diversity (interspecific interactions) on each of the denominators,  
402 total N yield and total DM yield (presented elsewhere for both the former (Grange et al.,  
403 unpubl.) and latter (Grange et al., in review)). Transgressive over yielding, whereby  
404 mixtures outperform the highest performing constituent monoculture, is driven by  
405 resource use efficiency and complementarity among species in mixtures (Mason et al.,  
406 2020).

407 Our study confirms that reduced emissions intensity can now be considered one of the  
408 many multi-functional benefits associated with multi-species swards. The 6-species  
409 mixture was more efficient, because more yield was produced with reductions in N<sub>2</sub>O  
410 losses to the environment. Overall, the six-species mixture significantly reduced N yield-  
411 scaled N<sub>2</sub>O emissions compared with *L. perenne* (both 150N and 300N) and 150N legume  
412 monocultures (Fig. 5a) and lower DM yield-scaled N<sub>2</sub>O emissions than 300N *L. perenne*  
413 (Fig. 5b). These results accord with the agronomic assessment of N<sub>2</sub>O emissions by van  
414 Groenigen et al (2010), where yield-scaled N<sub>2</sub>O emissions of non-leguminous crops  
415 increased rapidly at higher N application levels (>190 kg N ha<sup>-1</sup> year<sup>-1</sup>). As a practical  
416 consequence, in comparison to 300N *L. perenne*, the same N yield or DM yield could  
417 have been produced with the 150N six-species mixture using half the amount of fertiliser

418 and reducing N<sub>2</sub>O losses by 63% and 58% respectively. Similarly, in comparison to 150N  
419 *L. perenne*, the same N yield or DM yield could have been produced with the six-species  
420 mixture while reducing N<sub>2</sub>O losses by 41% and 24% respectively.

## 421 **5. Conclusion**

422 Overall, the effect of plant diversity on N<sub>2</sub>O emissions was derived from linear  
423 combinations of the species' performance in monoculture (species' identity). The effects  
424 of multi-species mixtures on N<sub>2</sub>O emissions intensity included species identity effects,  
425 and a net interspecific interaction that suppressed emissions intensity. The conventional  
426 300N *L. perenne* community produced over double the N<sub>2</sub>O emissions as the 150N six-  
427 species mixture (3.18 vs 1.52 kg N<sub>2</sub>O-N ha<sup>-1</sup> year<sup>-1</sup>). Considering emissions intensity, the  
428 same N yield and DM yield of 300N *L. perenne* could have been produced with the 6-  
429 species mixture using half the fertiliser and reduced N<sub>2</sub>O losses of 63% and 58%  
430 respectively. In comparison to 150N *L. perenne*, the same N yield and DM yield could  
431 have been produced with a 6-species mixture while producing 41% and 24% less N<sub>2</sub>O  
432 emissions. Communities dominated by legumes significantly increased N<sub>2</sub>O emissions,  
433 this should be considered when using legumes as cover crops. Overall, the manipulation  
434 of grassland composition is a practical, farm-scale management action that can reduce  
435 both N<sub>2</sub>O emissions and yield-scaled N<sub>2</sub>O emissions, and contribute to the sustainability  
436 of grassland production.

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449 **7. Appendices**

450 **Appendix A** Seeding rates of the multi-species experimental field trial. Species include *L. perenne* (Lp) *P.*  
 451 *pratense* (Pp), *T. pratense* (Tr), *T. repens* (Tr), *P. lanceolata* (Pl), and *C. intybus* (Ci).

Species	Lp	Pp	Tp	Tr	Pl	Ci
Seed (Kg ha <sup>-1</sup> )	28	12	12	15	10	8

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453 **Appendix B** Experimental design indicating the composition and relative abundance of the sown  
 454 communities. Also indicated are functional group richness (FGs), species richness (Species) and number of  
 455 replicates (Reps). Species include *L. perenne* (Lp) *P. pratense* (Pp), *T. pratense* (Tr), *T. repens* (Tr), *P.*  
 456 *lanceolata* (Pl), and *C. intybus* (Ci).

Community	Reps	FGs	Species	FG(G)	FG(H)	FG(L)	Lp	Pp	Tp	Tr	Pl	Ci
1	3	1	1	1	0	0	1	0	0	0	0	0
2	3	1	1	1	0	0	0	1	0	0	0	0
3	3	1	1	0	1	0	0	0	1	0	0	0
4	3	1	1	0	1	0	0	0	0	1	0	0
5	3	1	1	0	0	1	0	0	0	0	1	0
6	3	1	1	0	0	1	0	0	0	0	0	1
7	2	1	2	1	0	0	0.5	0.5	0	0	0	0
8	2	1	2	0	1	0	0	0	0.5	0.5	0	0
9	2	1	2	0	0	1	0	0	0	0	0.5	0.5
10	2	2	4	0.5	0.5	0	0.25	0.25	0.25	0.25	0	0
11	2	2	4	0.5	0	0.5	0.25	0.25	0	0	0.25	0.25
12	2	2	4	0	0.5	0.5	0	0	0.25	0.25	0.25	0.25
13	1	3	5	0.6	0.2	0.2	0.6	0	0.1	0.1	0.1	0.1
14	1	3	5	0.6	0.2	0.2	0	0.6	0.1	0.1	0.1	0.1
15	1	3	5	0.2	0.6	0.2	0.1	0.1	0.6	0	0.1	0.1
16	1	3	5	0.2	0.6	0.2	0.1	0.1	0	0.6	0.1	0.1
17	1	3	5	0.2	0.2	0.6	0.1	0.1	0.1	0.1	0.6	0
18	1	3	5	0.2	0.2	0.6	0.1	0.1	0.1	0.1	0	0.6
19	3	3	6	0.33	0.33	0.33	0.17	0.17	0.17	0.17	0.17	0.17
20	4	1	1	1	0	0	1	0	0	0	0	0

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461 **Appendix C** Fertiliser application rate equivalents to each plot over the agronomic year. (The community  
462 numbers are as listed in Appendix B.)

<b>Split</b>	<b>Date</b>	<b>Fertiliser application: communities 1-19</b>	<b>Fertiliser application: community 20</b>
1	12-Mar-2018	30 kg N ha <sup>-1</sup>	60 kg N ha <sup>-1</sup>
2	09-Apr-2018	30 kg N ha <sup>-1</sup>	60 kg N ha <sup>-1</sup>
3	09-May-2018	30 kg N ha <sup>-1</sup>	60 kg N ha <sup>-1</sup>
4	11-Jun-2018	20 kg N ha <sup>-1</sup>	40 kg N ha <sup>-1</sup>
5	20-Aug-2018	40 kg N ha <sup>-1</sup>	80 kg N ha <sup>-1</sup>

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464 **Appendix D** Aerial photograph of the experimental plot layout.



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640 **Figure legends**

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642 **Fig. 1.** Precipitation (mm) and temperature (°C) meteorological data for the experimental  
643 site collected from the JC meteorological station. Graph includes 3 x 30mm irrigation  
644 events which took place during summer 2018 of (due to drought).

645 **Fig. 2.** Nitrous oxide emissions and corresponding water filled pore space (WFPS %) of  
646 the experimental site for each sampling occasion. Emissions are displayed for the 6-  
647 species mixture, monocultures of the individual species (*L. perenne*, *P. pratense*, *T.*  
648 *pratense*, *T. repens*, *C. intybus* and *P. lanceolate*) and the 300N *L. perenne* monoculture.  
649 All communities received 150 kg ha<sup>-1</sup> year<sup>-1</sup> nitrogen fertiliser, except for the 300N *L.*  
650 *perenne* community that received 300 kg ha<sup>-1</sup> year<sup>-1</sup> of inorganic nitrogen fertiliser.  
651 Arrows indicate fertiliser application dates (*Appendix C*).

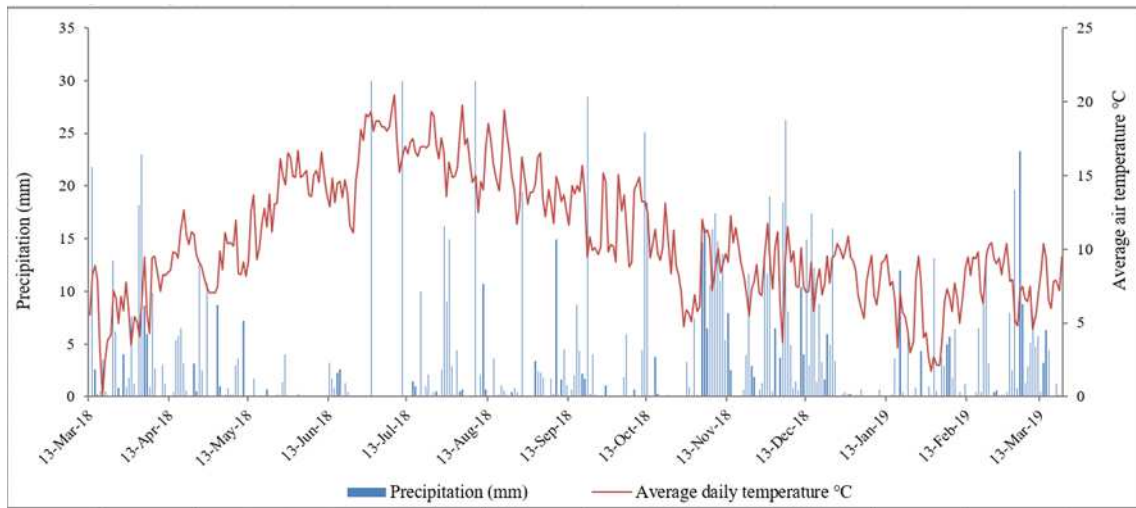
652 **Fig. 3.** Comparison of predicted N<sub>2</sub>O-N emissions (kg ha<sup>-1</sup> year<sup>-1</sup>) from monocultures  
653 and the 6-species mixture. Values that share the same letter are not significantly  
654 different ( $\alpha = 0.05$ ).

655 **Fig. 4 a)** Predicted annual N<sub>2</sub>O emissions (N<sub>2</sub>O-N kg ha<sup>-1</sup> year<sup>-1</sup>) in response to variation  
656 in the proportion of the grass (G), herb (H) and legume (L) FGs within grassland  
657 communities. The communities represented in this ternary diagram are based on the equi-  
658 proportional contribution of each of the two species within a FG. Thus, each vertex  
659 represents a 50:50 mixture of the two component species in the respective FG; the sides  
660 represent communities with varying proportions of two FGs (comprising four species),  
661 and the interior points represent varying proportions of three FGs (comprising six  
662 species). Thus, for example, the predicted N<sub>2</sub>O emissions for the community comprising

663 10% grass, 40% legume and 50% herb is calculated from the species-level composition  
664 comprising 5% *L. perenne*, 5% *P. pratense*, 20% *T. pratense*, 20% *T. repens*, 25% *C.*  
665 *intybus* and 25% *P. lanceolata*. **b**) Predicted annual N<sub>2</sub>O emissions (N<sub>2</sub>O-N kg ha<sup>-1</sup> year<sup>-1</sup>)  
666 for a 50:50 mixture of the two species from each FG: grass, legume and herb (these  
667 predictions correspond to the vertices in the ternary diagram). For example, the legume  
668 FG contains 50% *T. pratense* and 50% *T. repens*. Bars that share a letter are not  
669 significantly different ( $\alpha = 0.05$ ).

670 **Fig. 5. a and b** Comparison of yield-scaled N<sub>2</sub>O emissions from forage monocultures,  
671 the 6-species mixture and the mean of the six 150N monocultures for **a**) N yield-scaled  
672 N<sub>2</sub>O emissions (N<sub>2</sub>O-N g ha<sup>-1</sup>year<sup>-1</sup>/N yield kg ha<sup>-1</sup> year<sup>-1</sup>) and **b**) DM yield-scaled N<sub>2</sub>O  
673 emissions (N<sub>2</sub>O-N g ha<sup>-1</sup> year<sup>-1</sup>/DM yield tonne ha<sup>-1</sup> year<sup>-1</sup>). Values that share the same  
674 letter are not significantly different ( $\alpha = 0.05$ ).

675 **Fig. 6.** Estimated emission intensity analyses for **a**) N yield-scaled N<sub>2</sub>O emissions (N<sub>2</sub>O  
676 g ha<sup>-1</sup> year<sup>-1</sup> / N yield kg ha<sup>-1</sup> year<sup>-1</sup>) and **b**) DM yield-scaled N<sub>2</sub>O emissions (N<sub>2</sub>O g ha<sup>-1</sup>  
677 year<sup>-1</sup>/DM yield tonne ha<sup>-1</sup> year<sup>-1</sup>) in response to variation in the proportion of grass (G),  
678 herb (H) and legume (L) FGs within grassland communities. The communities  
679 represented in this ternary diagram are based on an equal proportional contribution of  
680 each of the two species within a FG. Thus, each vertex indicates the average of the two  
681 component species in the respective FG; the sides represent communities with varying  
682 proportions of two FGs (comprising four species), and the interior points represent  
683 varying proportions of three FGs (comprising six species).

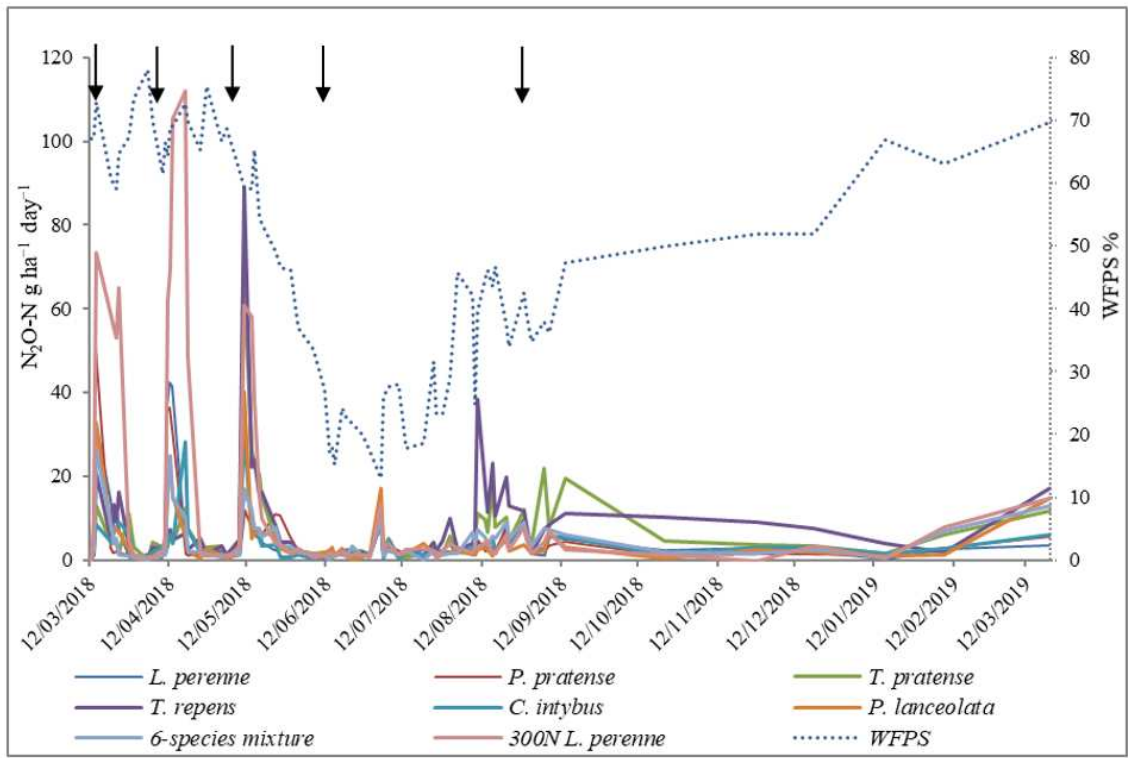


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686 Fig. 1

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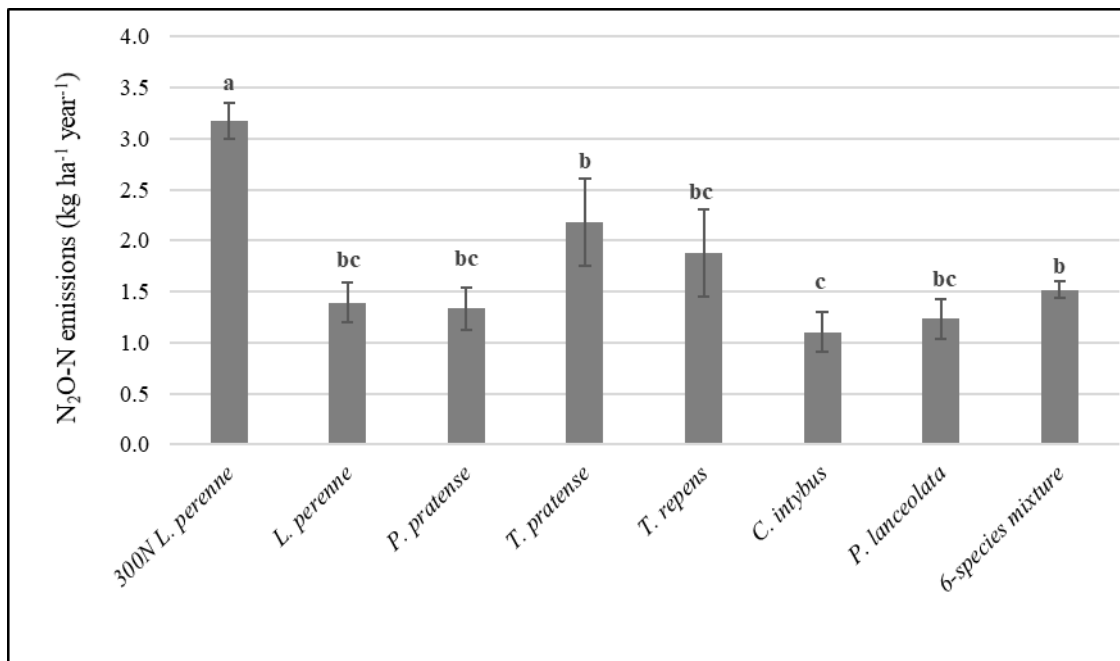
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690 Fig. 2

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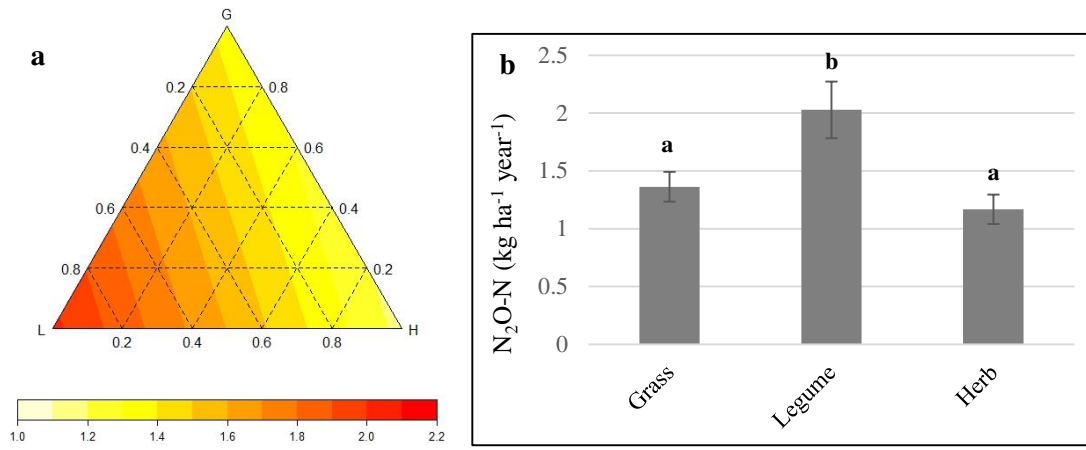
694 **Fig. 3**

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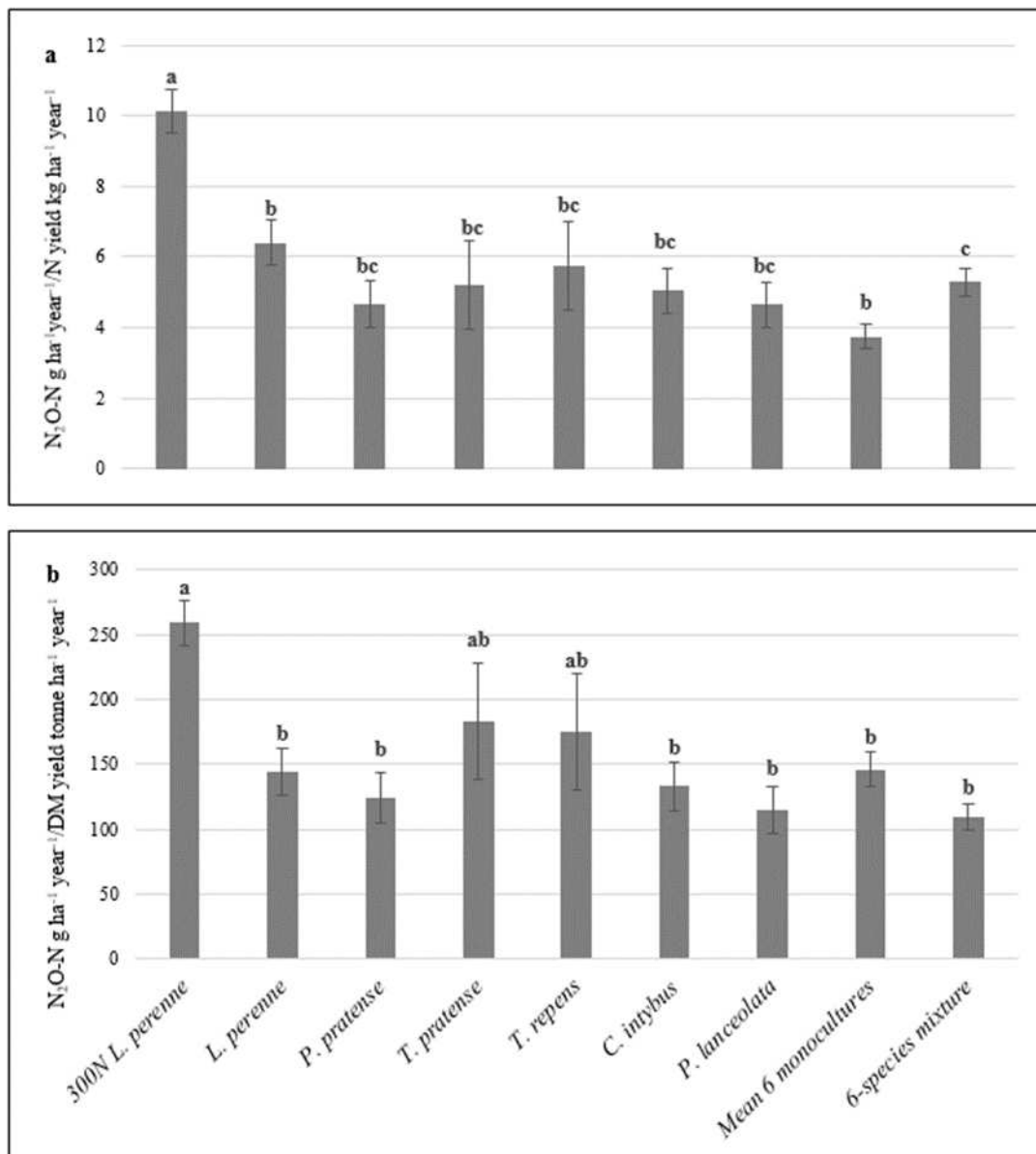


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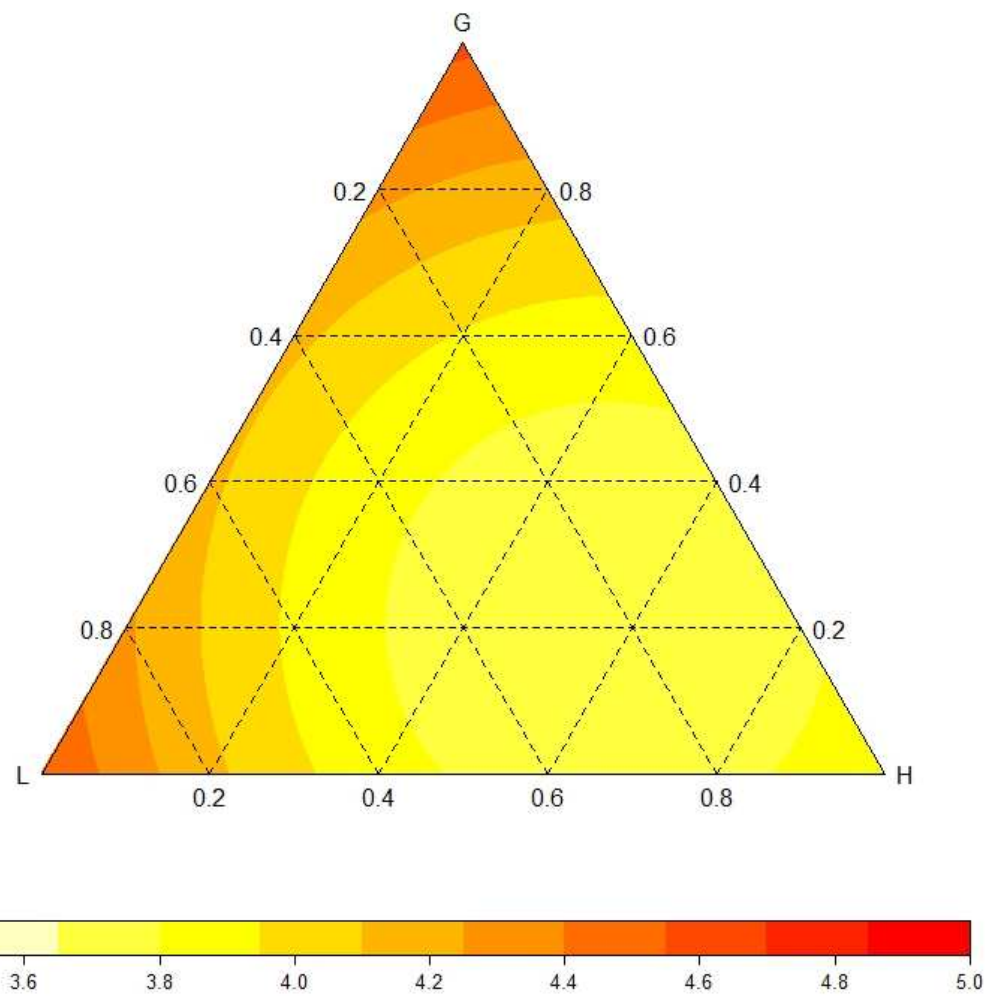
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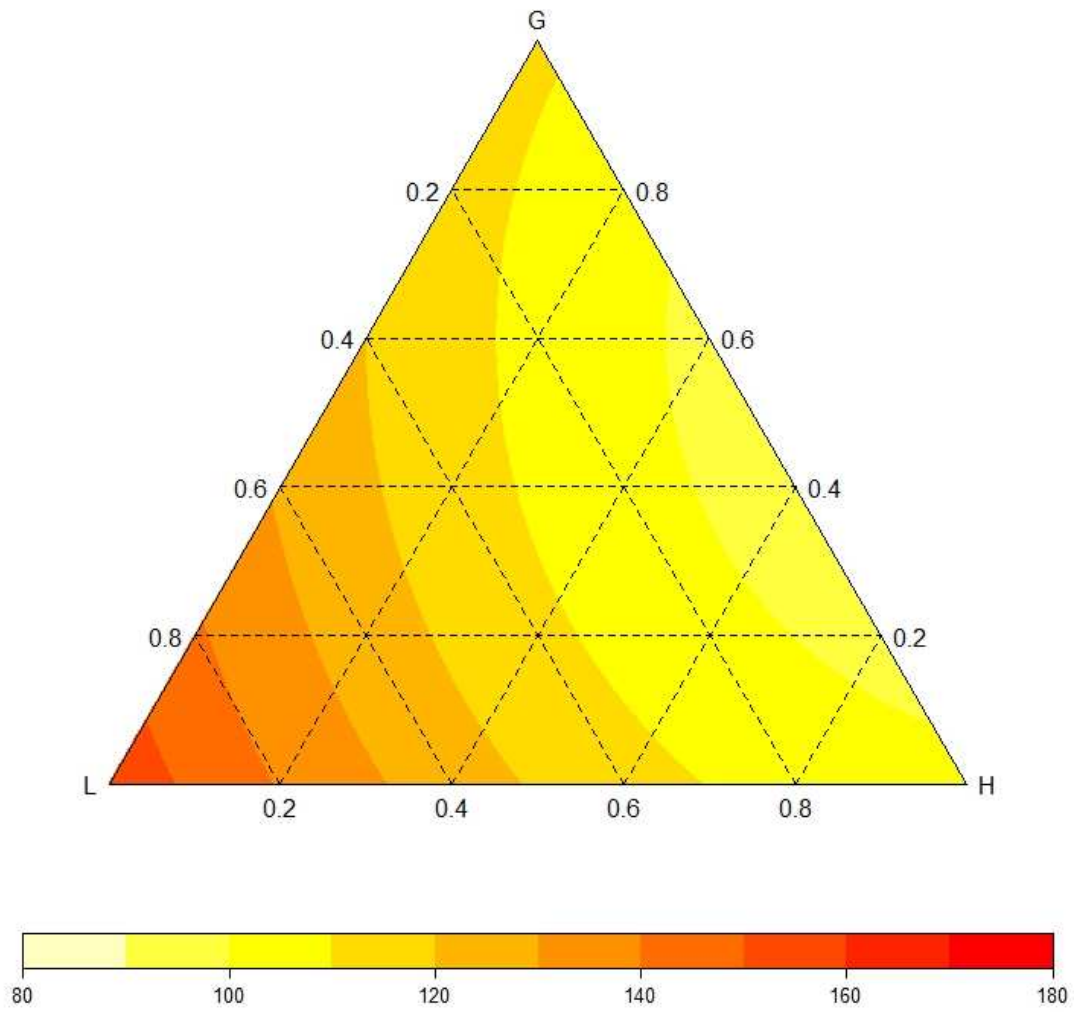
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726 **Fig. 6b**

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**Table 1.** (a) Coefficient estimates  $\pm$  standard errors for the identity effects ( $\beta$ ) and interaction estimates ( $\delta$ ) from equation 2, and (b) predictions for the average monoculture and the equi-proportional 6-species mixtures. These are presented for the models fitted to each of the three responses: N<sub>2</sub>O-N emissions, N yield-scaled N<sub>2</sub>O emissions and DM yield-scaled N<sub>2</sub>O emissions.

Modelled estimates				
		N <sub>2</sub> O emissions	N yield-scaled N <sub>2</sub> O emissions	DM yield-scaled N <sub>2</sub> O emissions
		(N <sub>2</sub> O-N kg ha <sup>-1</sup> year <sup>-1</sup> )	(N <sub>2</sub> O-N g ha <sup>-1</sup> year <sup>-1</sup> /N yield kg ha <sup>-1</sup> year <sup>-1</sup> )	(N <sub>2</sub> O-N g ha <sup>-1</sup> year <sup>-1</sup> /DM yield tonne ha <sup>-1</sup> year <sup>-1</sup> )
(a)	300N <i>L. perenne</i>	3.18 $\pm$ 0.196	10.14 $\pm$ 0.603	259.3 $\pm$ 17.46
	150N <i>L. perenne</i>	1.39 $\pm$ 0.198	6.39 $\pm$ 0.644	144.4 $\pm$ 18.68
	<i>P. pratense</i>	1.33 $\pm$ 0.206	4.65 $\pm$ 0.659	124.1 $\pm$ 19.11
	<i>T. pratense</i>	2.18 $\pm$ 0.428	5.21 $\pm$ 1.269	183.6 $\pm$ 44.95
	<i>T. repens</i>	1.87 $\pm$ 0.428	5.76 $\pm$ 1.269	174.6 $\pm$ 44.95
	<i>C. intybus</i>	1.10 $\pm$ 0.197	5.04 $\pm$ 0.640	133.2 $\pm$ 18.57
	<i>P. lanceolata</i>	1.23 $\pm$ 0.197	4.66 $\pm$ 0.640	115.0 $\pm$ 18.57
	Species interaction effect $\delta$	n/a	-3.69 $\pm$ 1.394	-87.1 $\pm$ 43.98
(b)	6-species mixture	1.52 $\pm$ 0.083	3.75 $\pm$ 0.356	109.5 $\pm$ 10.33
	Mean of 6 monocultures	1.52 $\pm$ 0.083	5.29 $\pm$ 0.401	145.8 $\pm$ 13.51

**Saoirse Cummins:** data curation (nitrous oxide), formal analysis, visualization, writing – original draft preparation.

**John A. Finn:** supervision, methodology, formal analysis, visualization, writing- reviewing and editing

**Karl G. Richards:** supervision, methodology, writing- reviewing and editing

**Gary J. Lanigan:** supervision, methodology, writing- reviewing and editing

**Guylain Grange:** data curation (yield)

**Caroline Brophy:** methodology, formal analysis

**Laura M. Cardenas:** methodology, writing- reviewing and editing

**Tom H. Misselbrook:** methodology, writing- reviewing and editing

**Chris K. Reynolds:** methodology, writing- reviewing and editing

**Dominika J. Krol:** supervision, methodology, writing- reviewing and editing

**Declaration of interests**

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

The authors declare the following financial interests/personal relationships which may be considered as potential competing interests: